



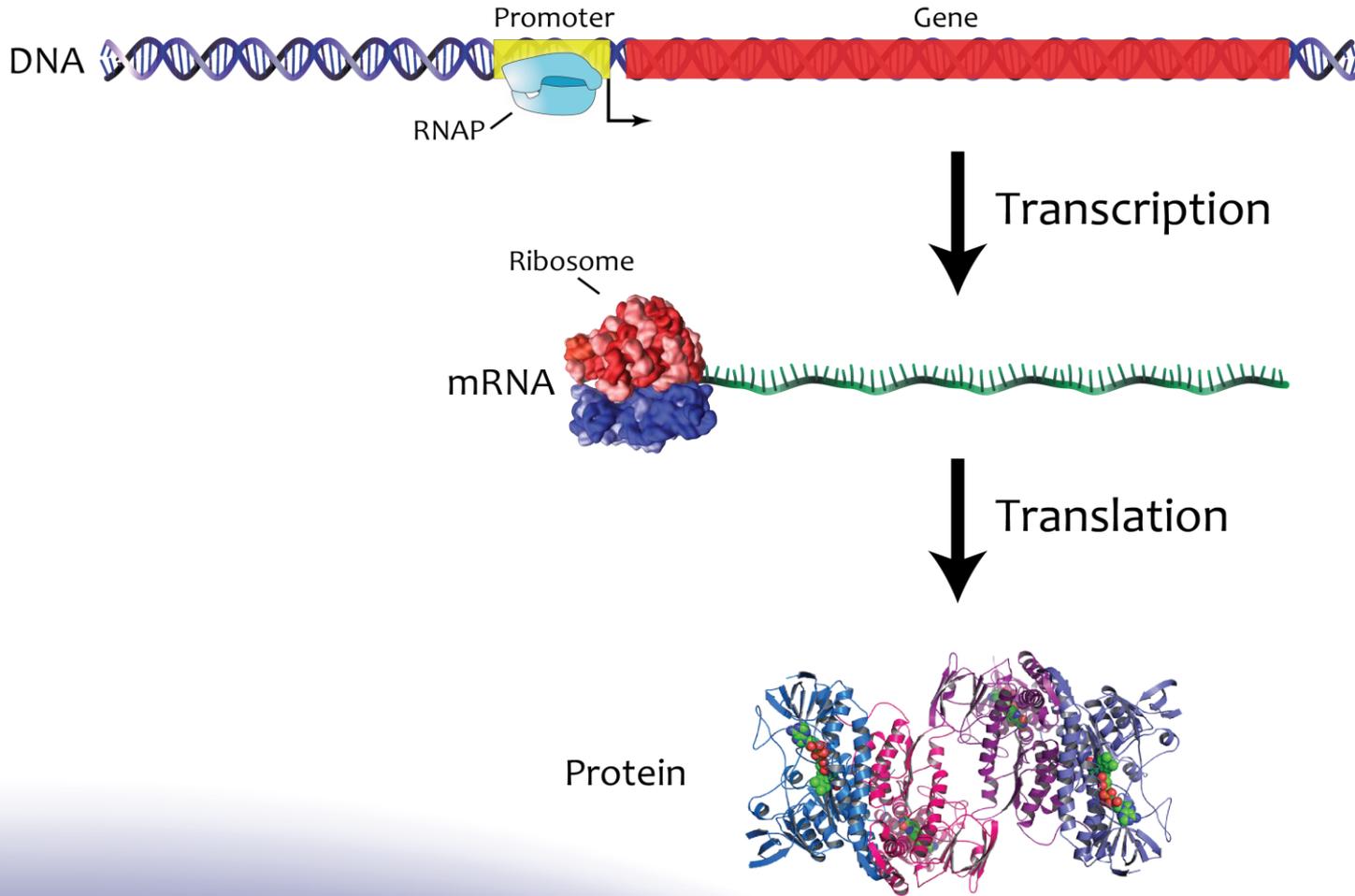
# Enhancer Modeling by Monte-Carlo Simulations

Yaroslav Pollak

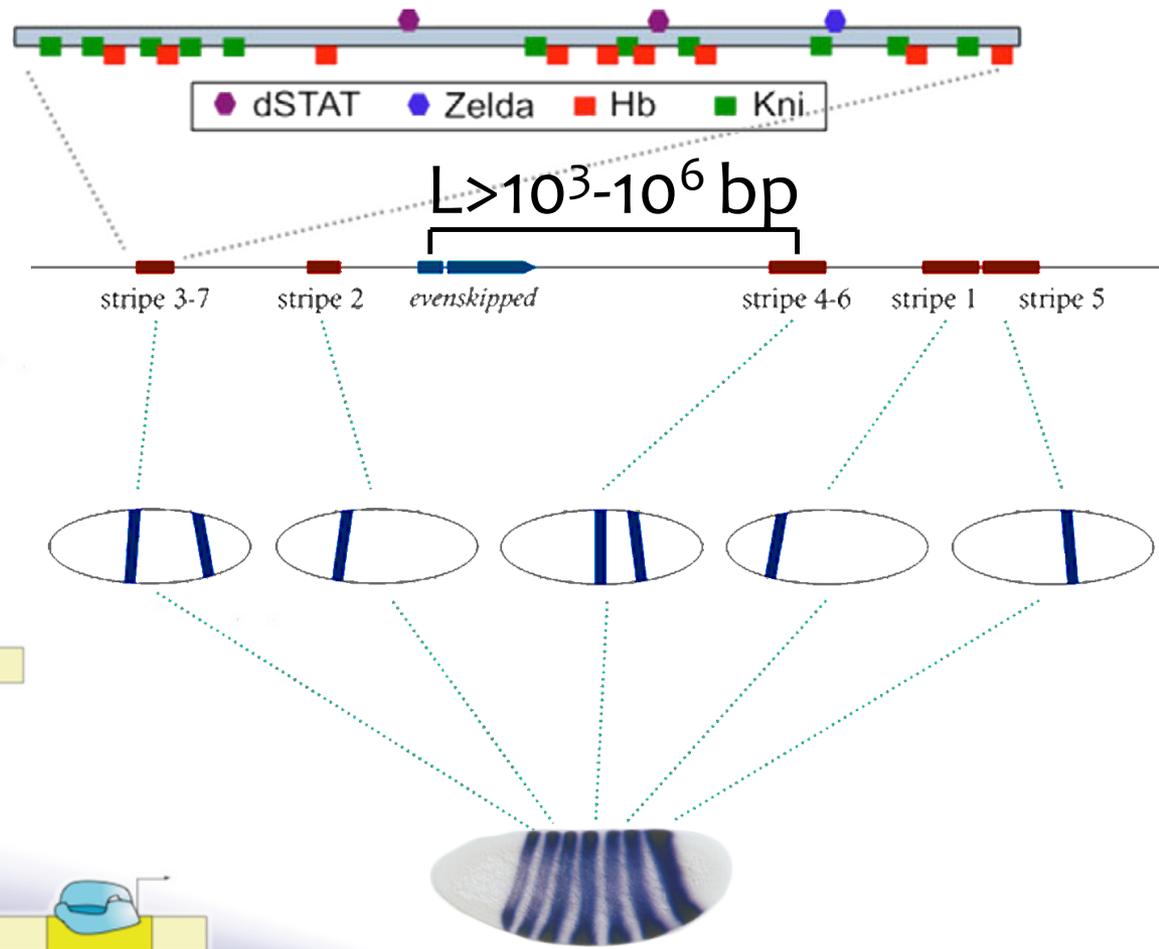
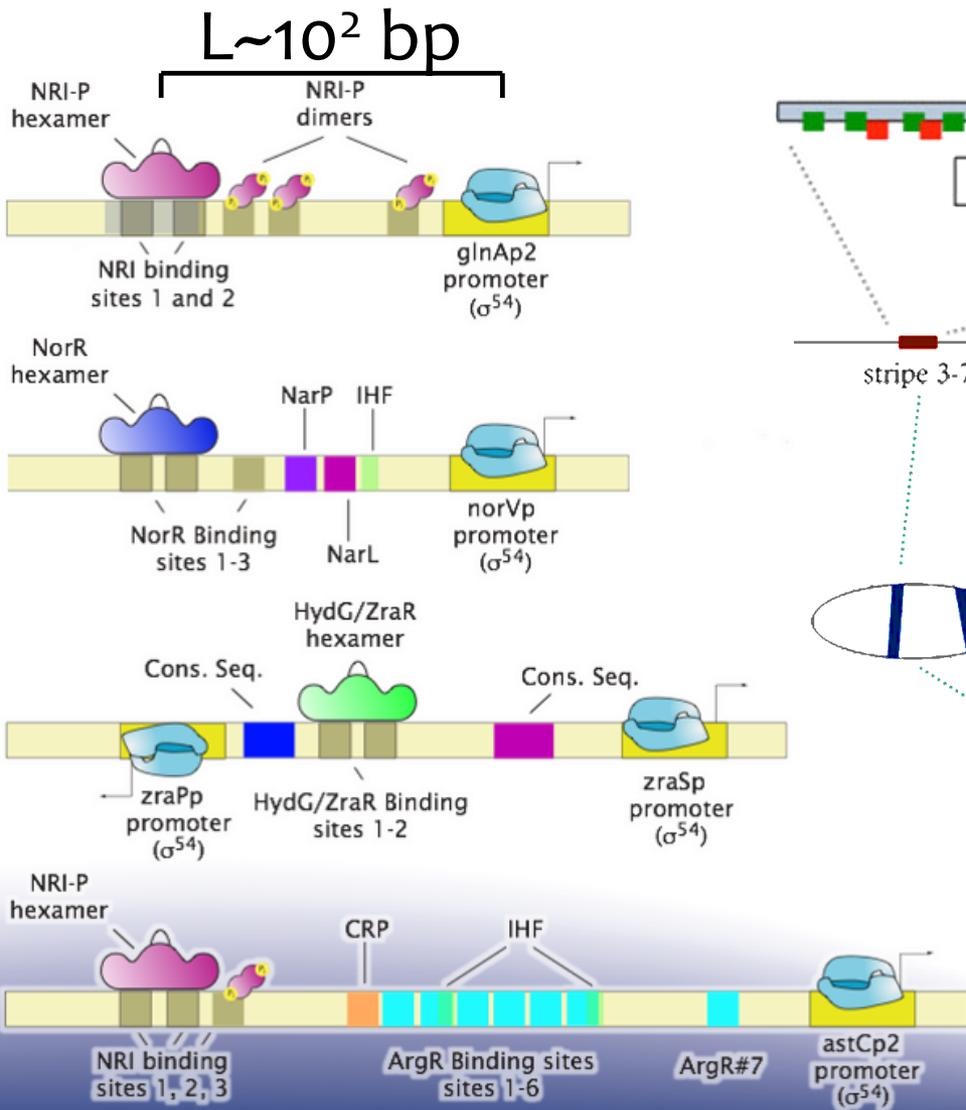
PhD seminar  
November, 2016

# Introduction

# Central Dogma of Biology

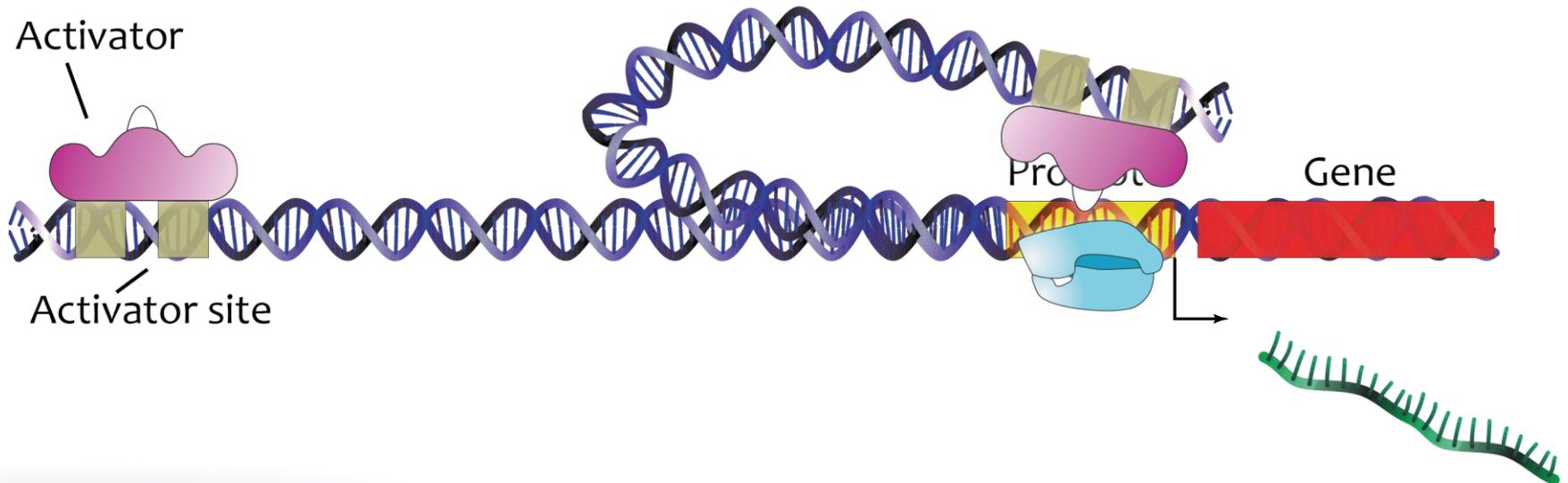


# Enhancers

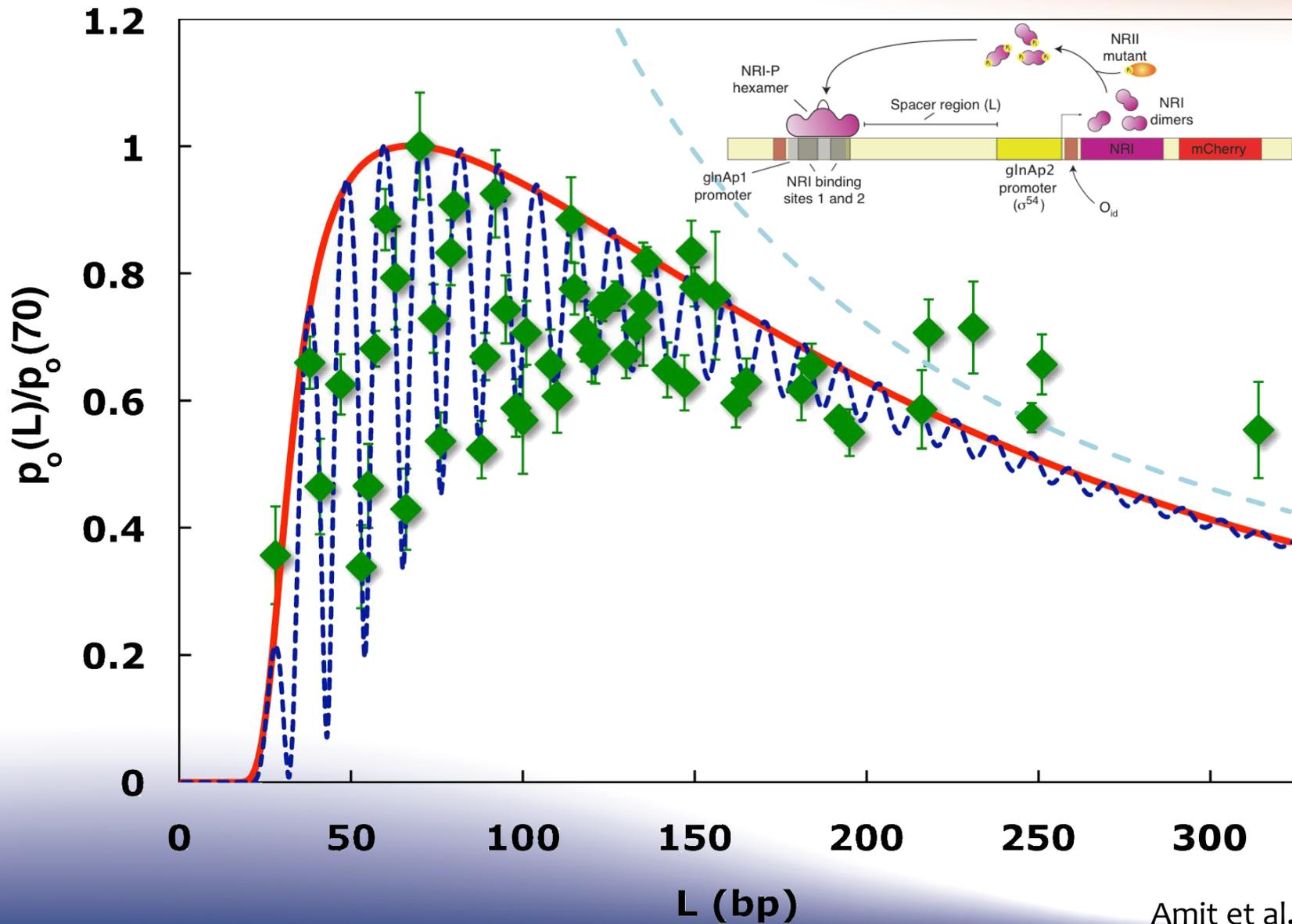


# Looping-based Transcription

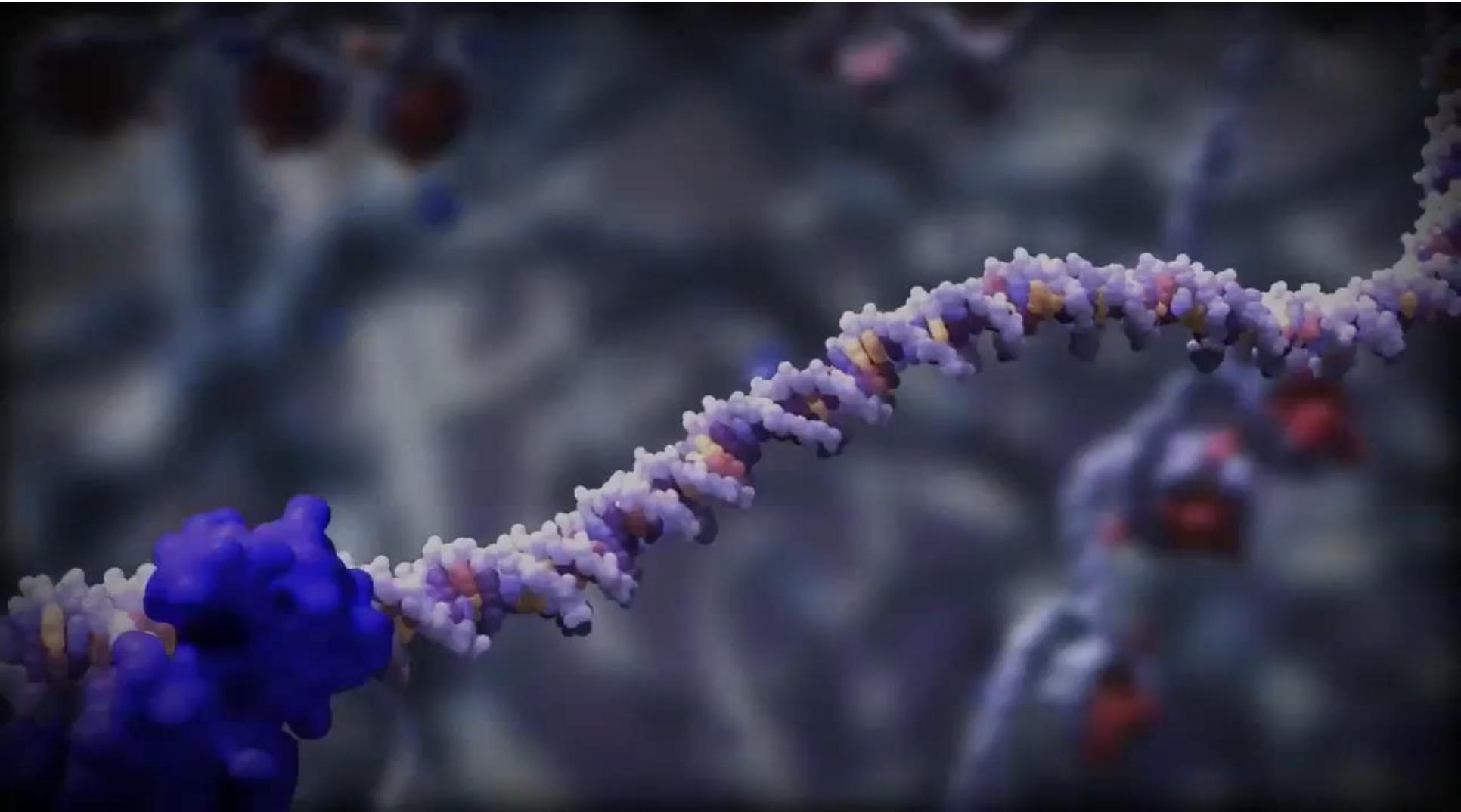
- DNA is constantly changing conformations
- Every once in a while it loops (stochastic process)
- Looping probability  $\Rightarrow$  Transcriptional activity



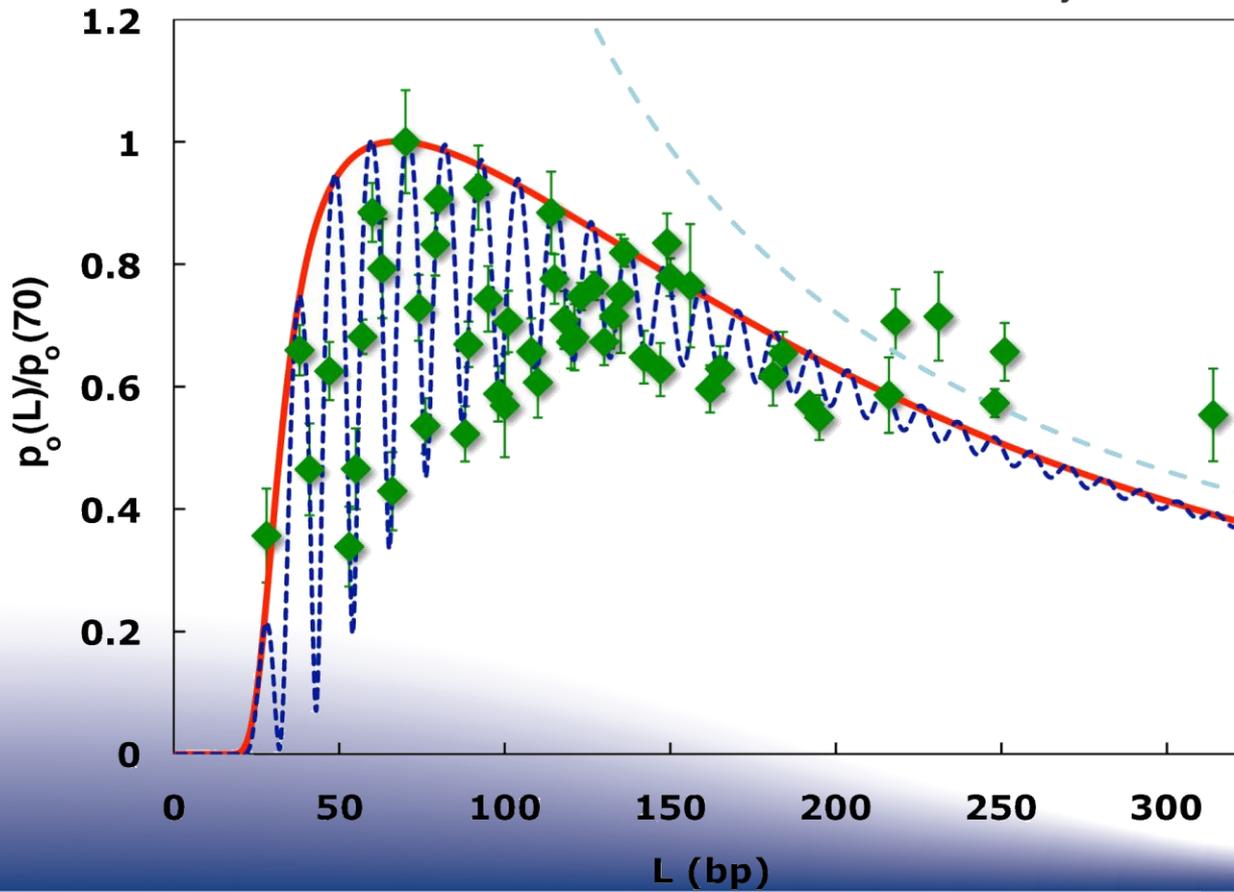
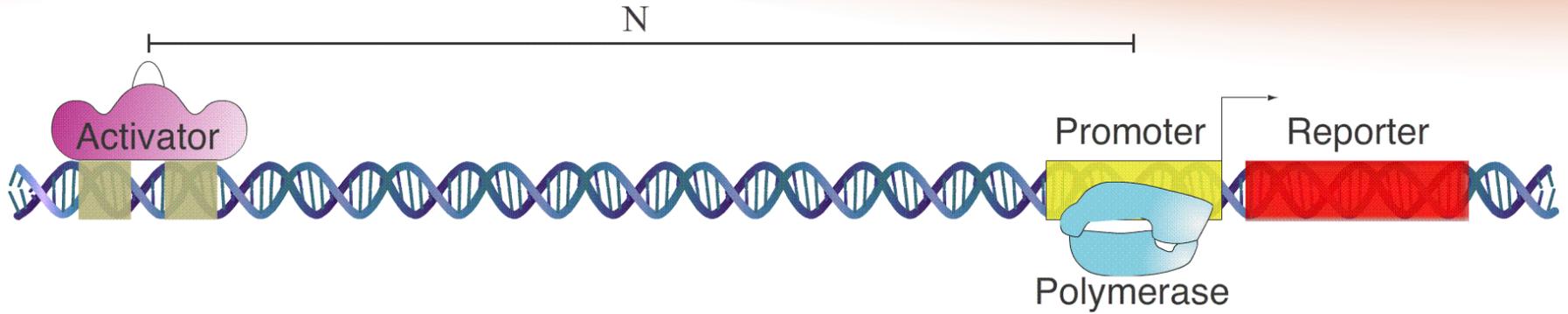
# Looping Shown in Bacteria



# Protein Binding to DNA

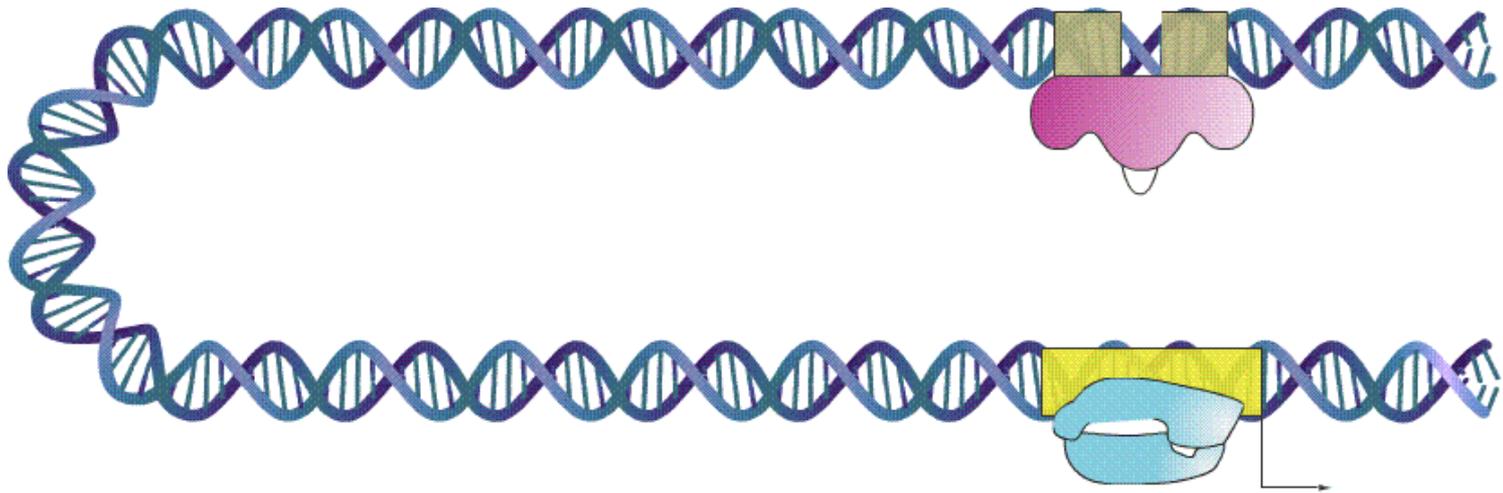


# Oscillations



# Transcription Regulation

- Regulation by Transcription Factors (TFs) binding DNA



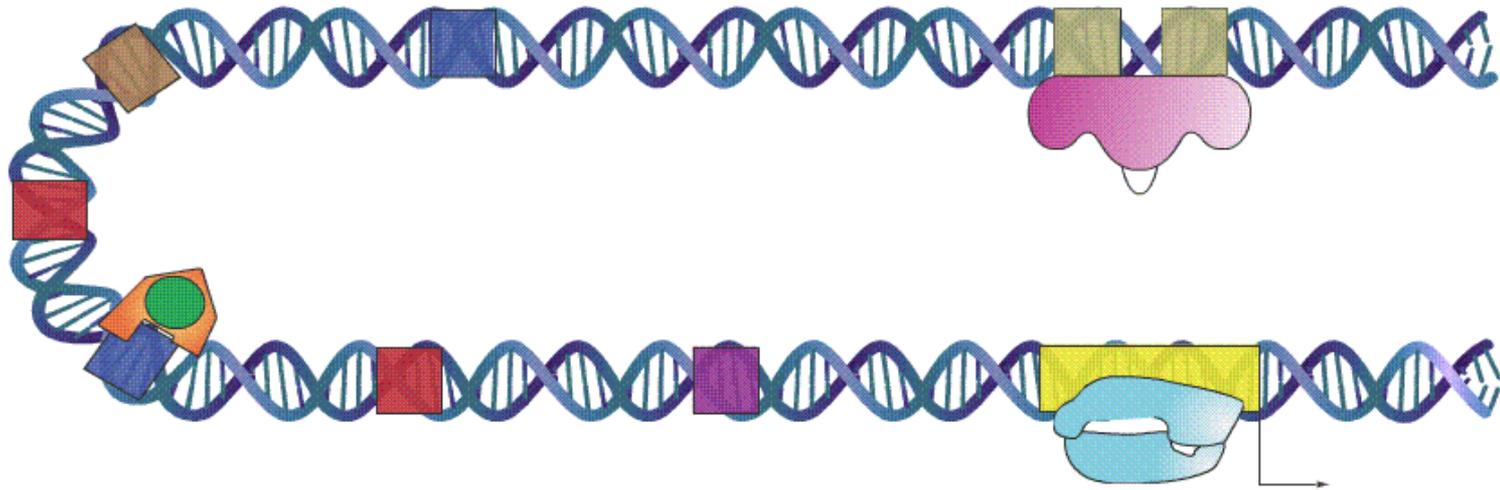
# Transcription Regulation

- Regulation by Transcription Factors (TFs) binding DNA



# Transcription Regulation

- Regulation by Transcription Factors (TFs) binding DNA

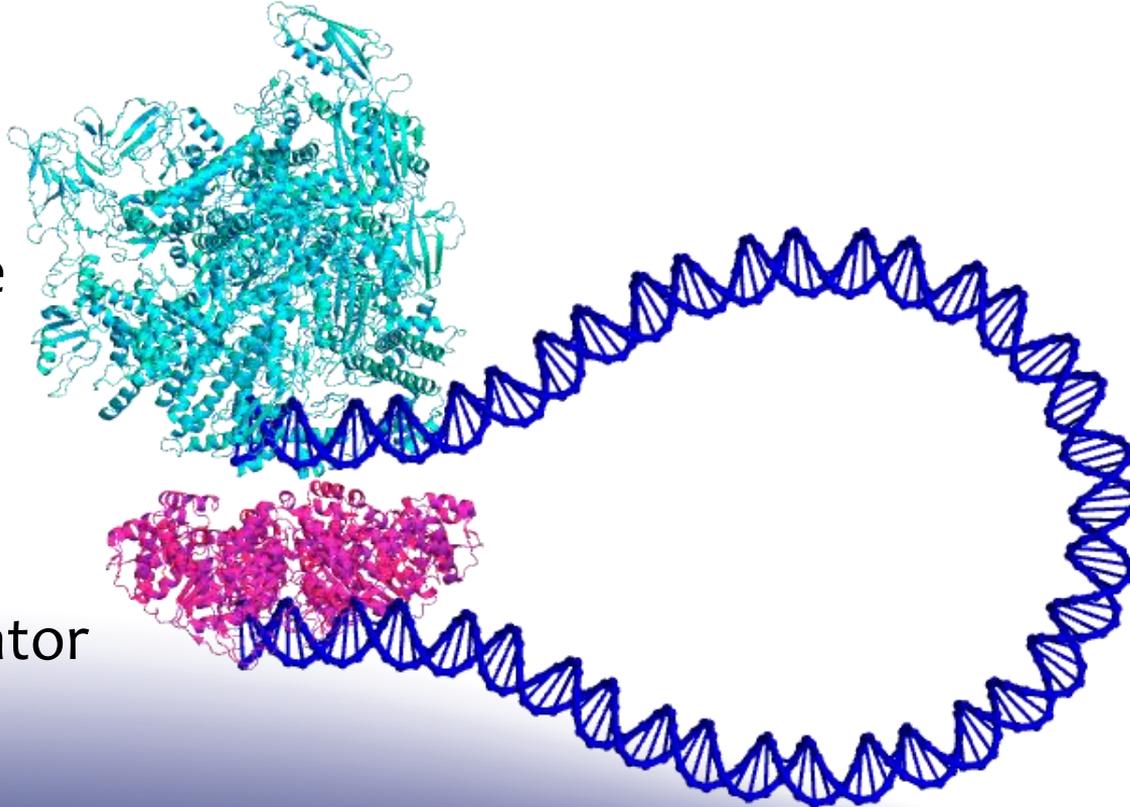


# Physical Mechanism

- Known mechanisms: TF alters DNA locally
  - Bending – No long-range effect, few TFs bend DNA
  - Twisting – No long-range effect, few TFs twist DNA
  - Stiffening – No long-range effect
- Not addressing chromatin modification

# Excluded Volume

RNA  
Polymerase

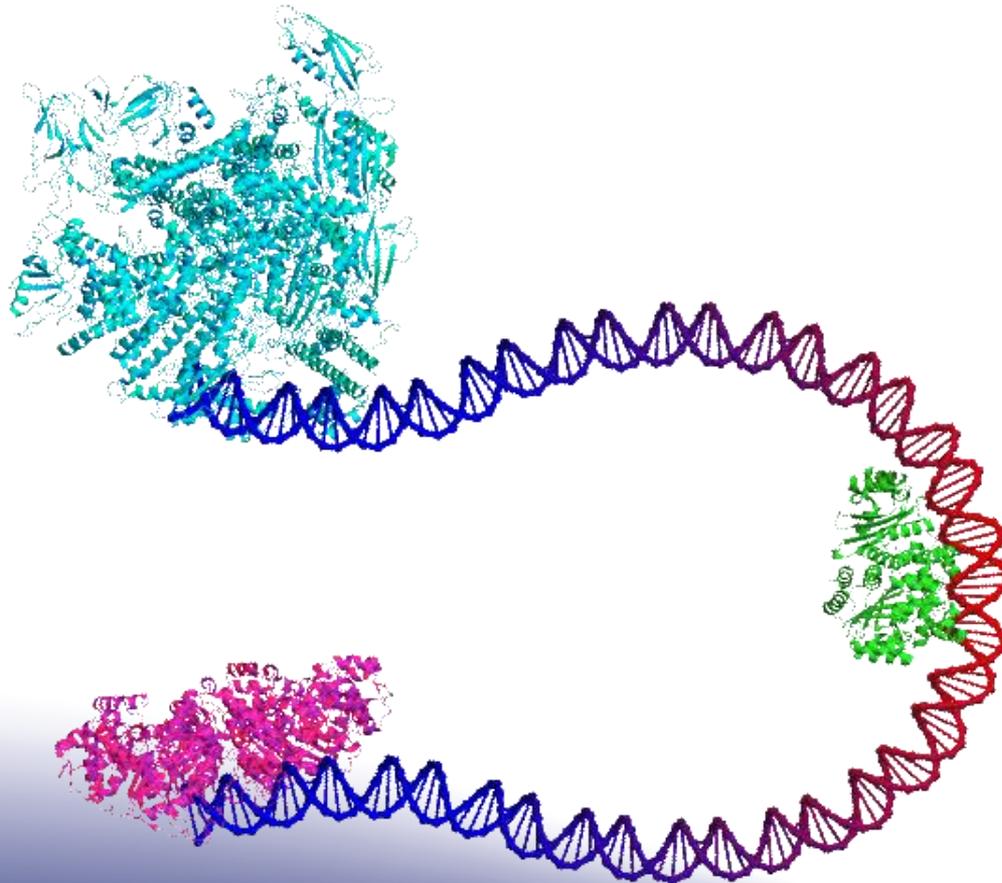


Activator

# Excluded Volume

- Any\* DNA binding protein can **increase** or **decrease** looping probability depending its orientation

RNA  
Polymerase

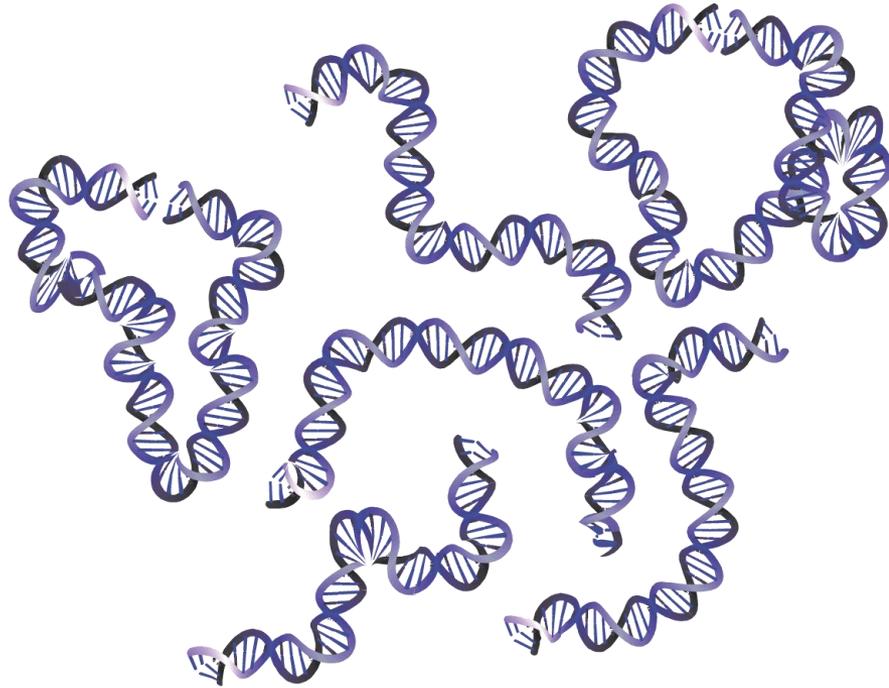


TF

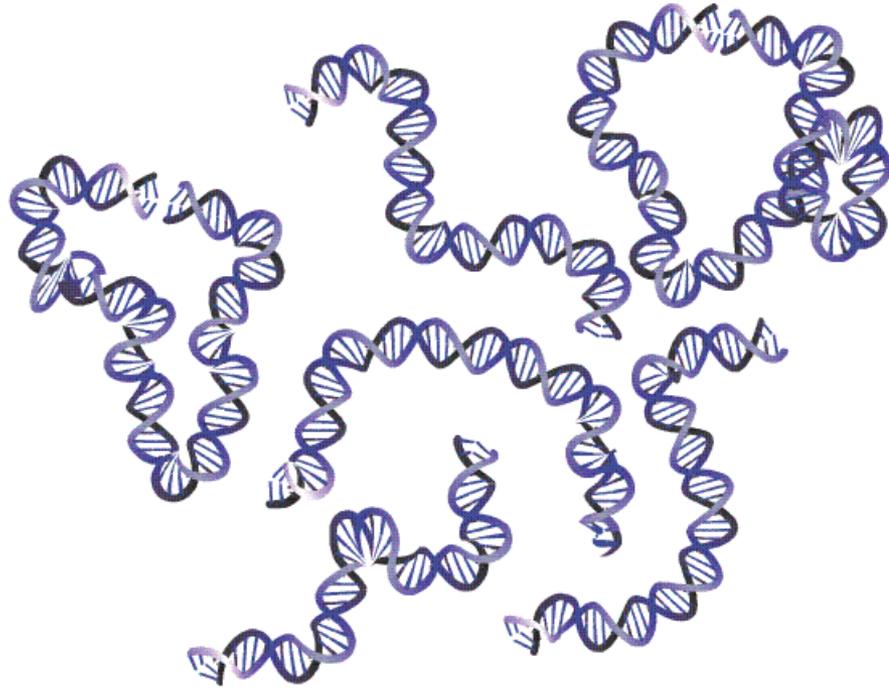
Activator

# Modeling

# Looping Probability



# Looping Probability



# Looping Probability

$$P_{loop} = \frac{e^{-\beta E_{loop}} + e^{-\beta E_{open}}}{e^{-\beta E_{loop}} + e^{-\beta E_{loop}}}$$

$\frac{1}{k_b T}$   
↓



# DNA Model



## DNA

Discrete chain of straight links. For link  $i$ :

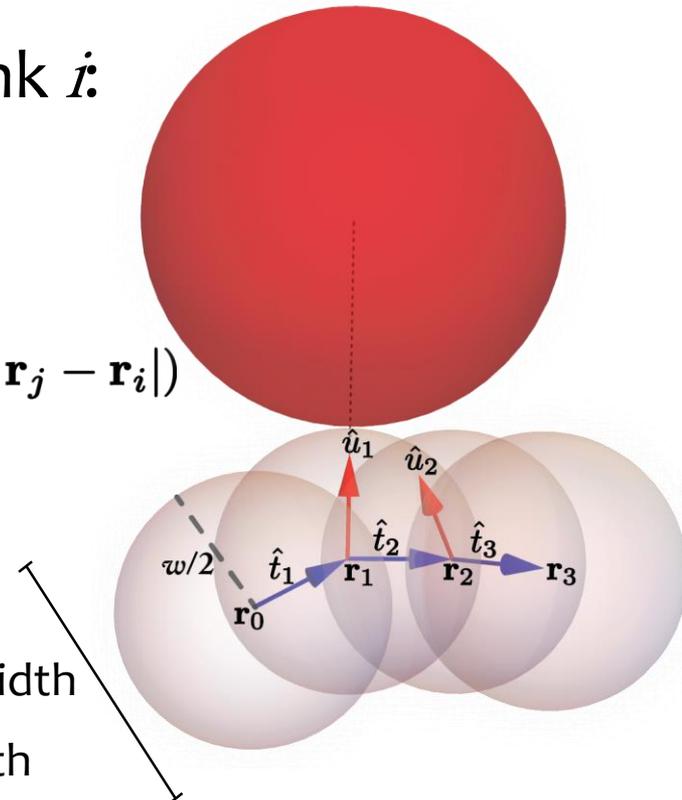
- Bending energy:  $\beta E_i^{bend} = \frac{a}{2} |\hat{t}_i - \hat{t}_{i-1}|^2$
- Twisting energy:  $\beta E_i^{twist} = c (\Omega_i - \Omega_0)^2$
- Chain volume:  $\beta E_i^{hw} = \infty \sum_{j \neq i} \Theta(w - |\mathbf{r}_j - \mathbf{r}_i|)$

## Regulator TF

- Local stiffening:  $a \rightarrow a'$
- Local bending:  $\hat{t}_i \rightarrow \hat{t}'_i$

## TFs & RNAP

- Spherical volume



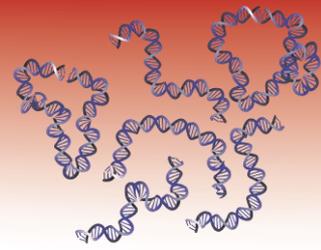
Effective width  
>  
DNA width

Wang et al., Macromolecules 2011

Pollak et al., PRE 2014

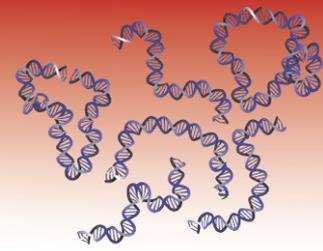
Brunwasser-Meirom, Pollak et al., Nat. Comm 2016

# Monte-Carlo Simulations

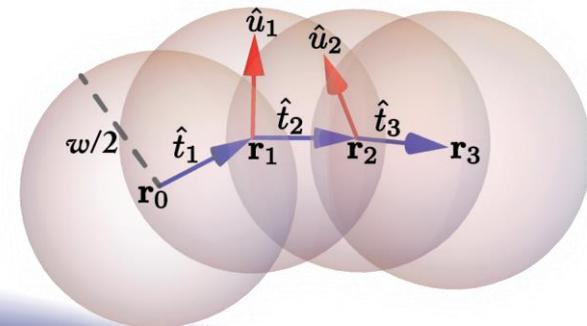
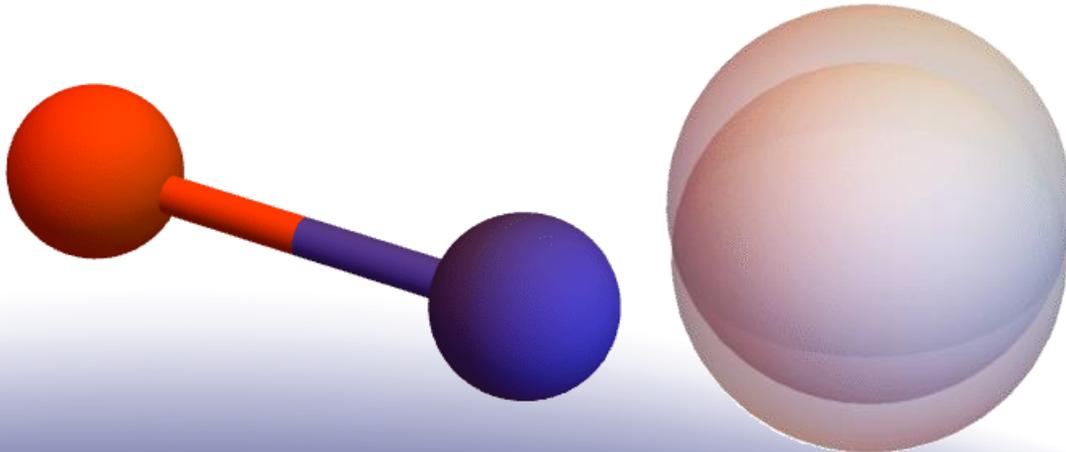


- Chain & TF volumes make considering all possible DNA conformations very difficult
- Static Monte-Carlo (Sequential Importance Sampling)

# Monte-Carlo Simulations



- Static Monte-Carlo (Sequential Importance Sampling)
- Chains generated from scratch, link after link
- Samples of  $\sim 10^9$  chains with & without TFs
- Comparing looping probability with & without TFs determines regulatory effect.



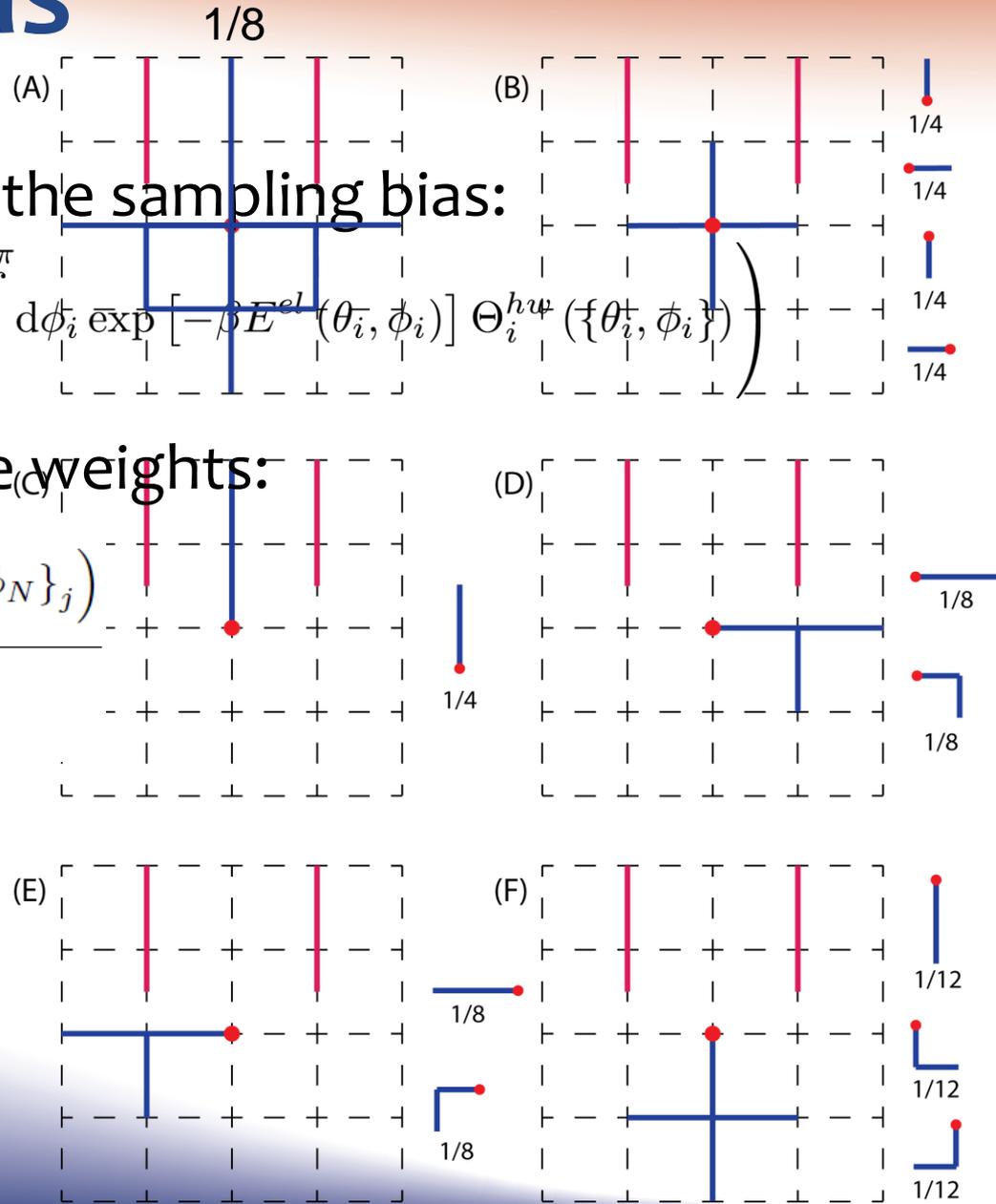
# Sampling Bias

- Need weights to offset the sampling bias:

$$W(\{\theta_N, \phi_N\}) = \prod_{i=2}^N \left( \int_{-1}^1 d \cos \theta_i \int_0^{2\pi} d\phi_i \exp[-\beta E^{el}(\theta_i, \phi_i)] \Theta_i^{hw}(\{\theta_i^+, \phi_i^+\}) \right)$$

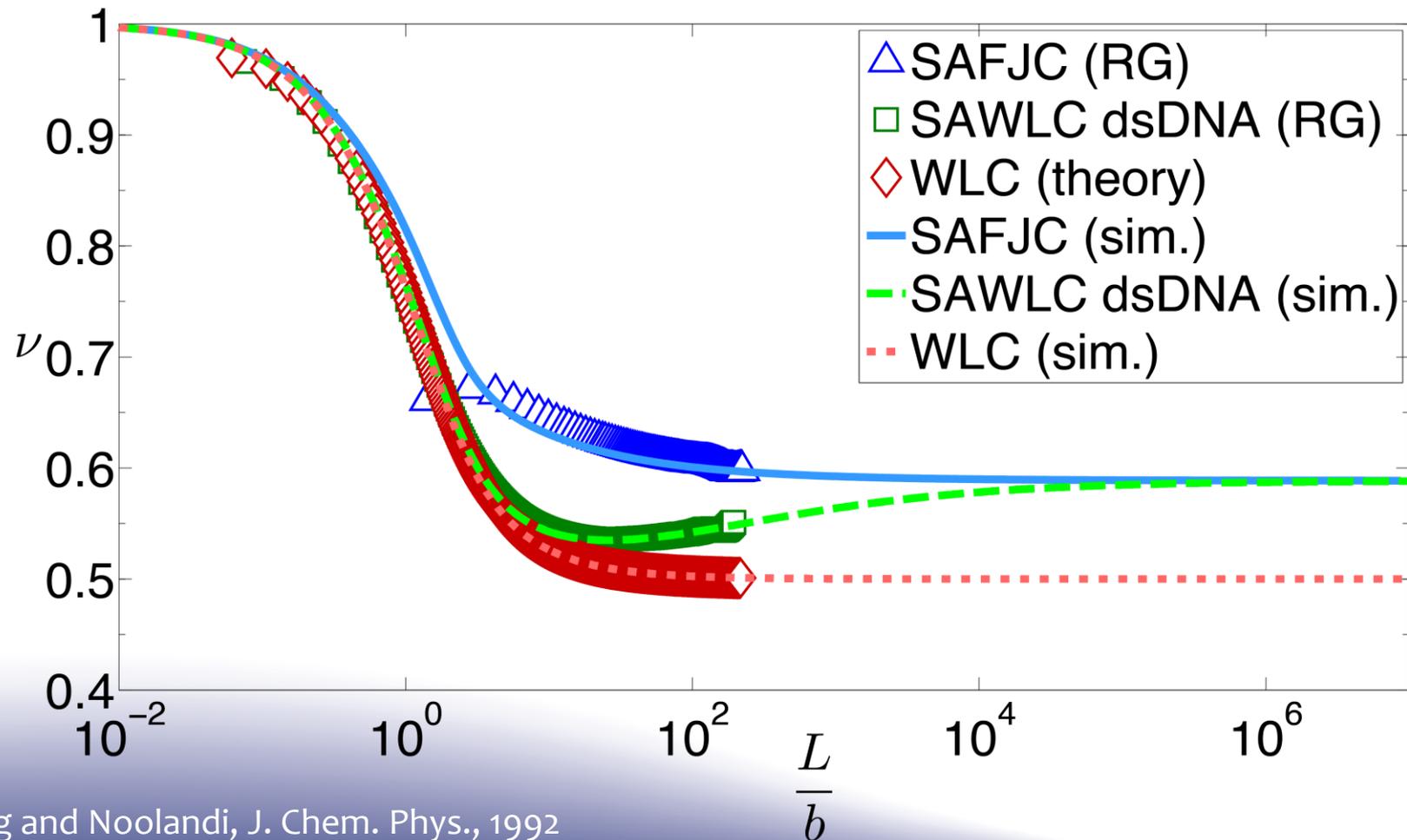
- Ensemble estimates use weights:

$$\langle f \rangle = \frac{\sum_{j=1}^{N_c} f(\{\theta_N, \phi_N\}_j) W(\{\theta_N, \phi_N\}_j)}{\sum_{j=1}^{N_c} W(\{\theta_N, \phi_N\}_j)}$$



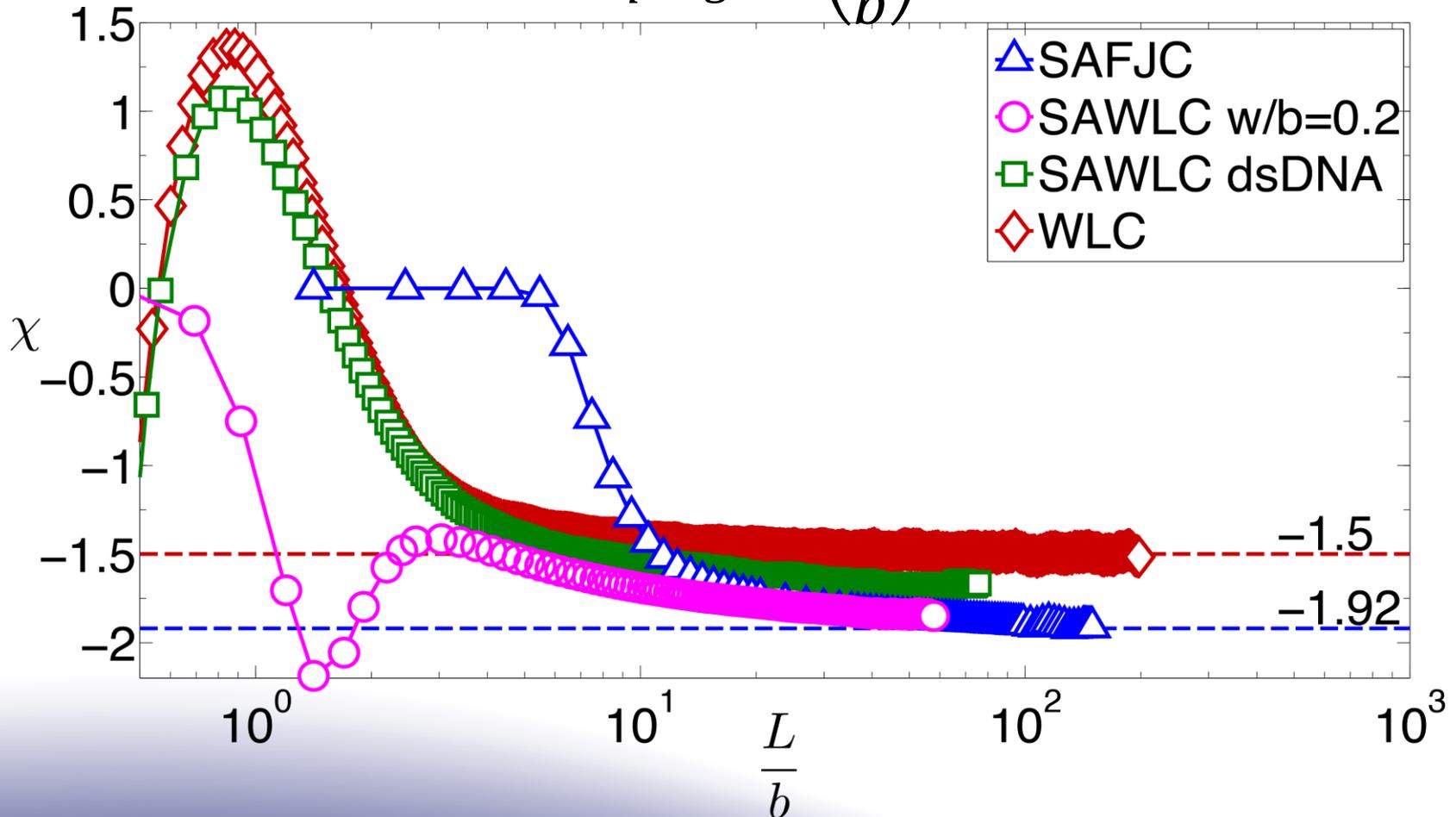
# DNA Model Verification

$$\sqrt{\langle R^2 \rangle} \propto \left(\frac{L}{b}\right)^\nu$$



# DNA Model Verification

$$P_{\text{looping}} \propto \left(\frac{L}{b}\right)^\chi$$



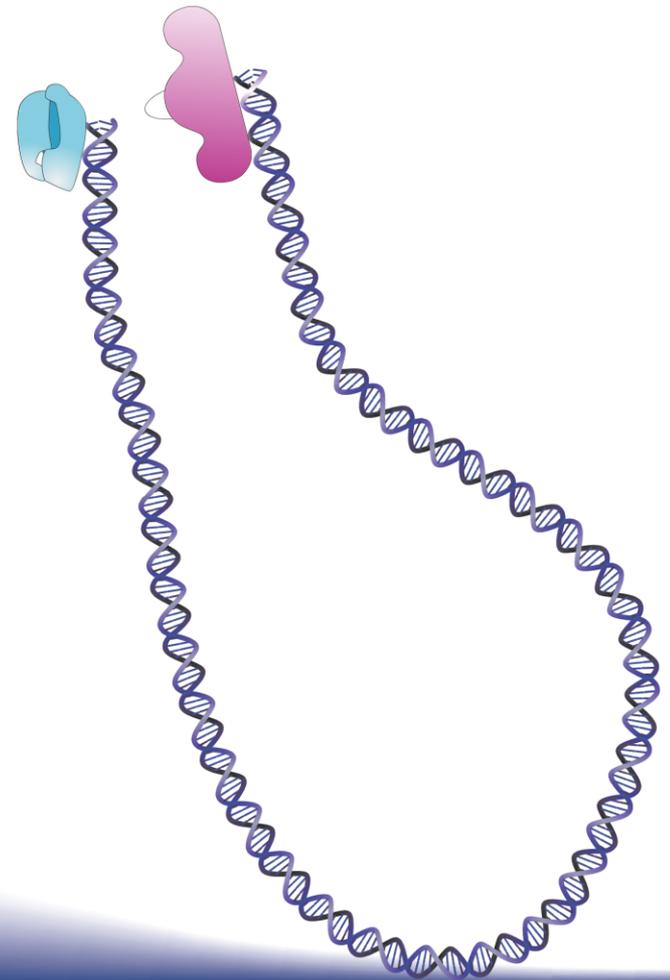
Genes, Cornell U. Press, 1979, Sinclair et al., JACS, 1985

Pollak et al., PRE 2014

# Short-Range Looping Results

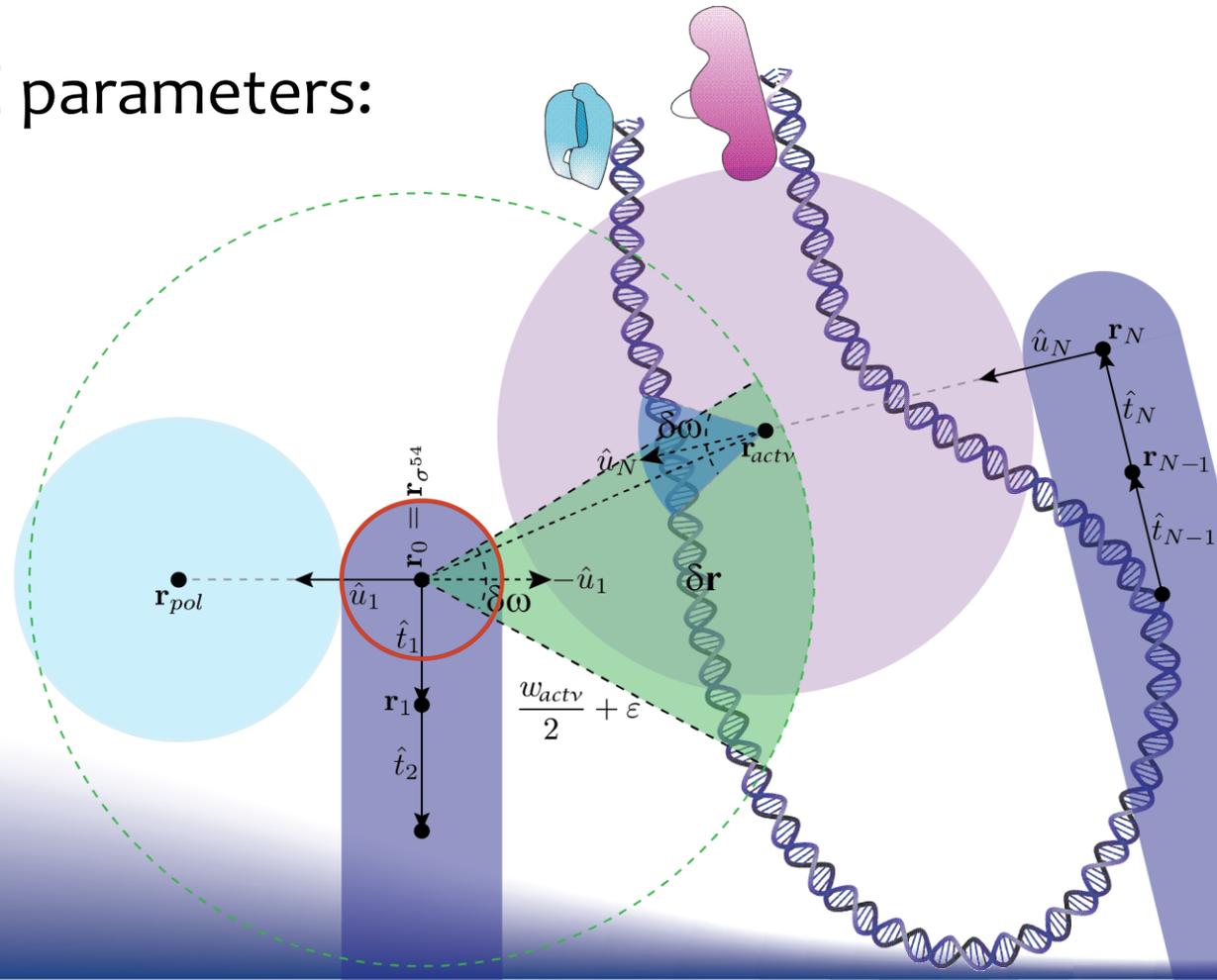
# Simulation Looping Condition

- Looping conditions mimicking bacterial  $\sigma^{54}$  promoters



# Simulation Looping Condition

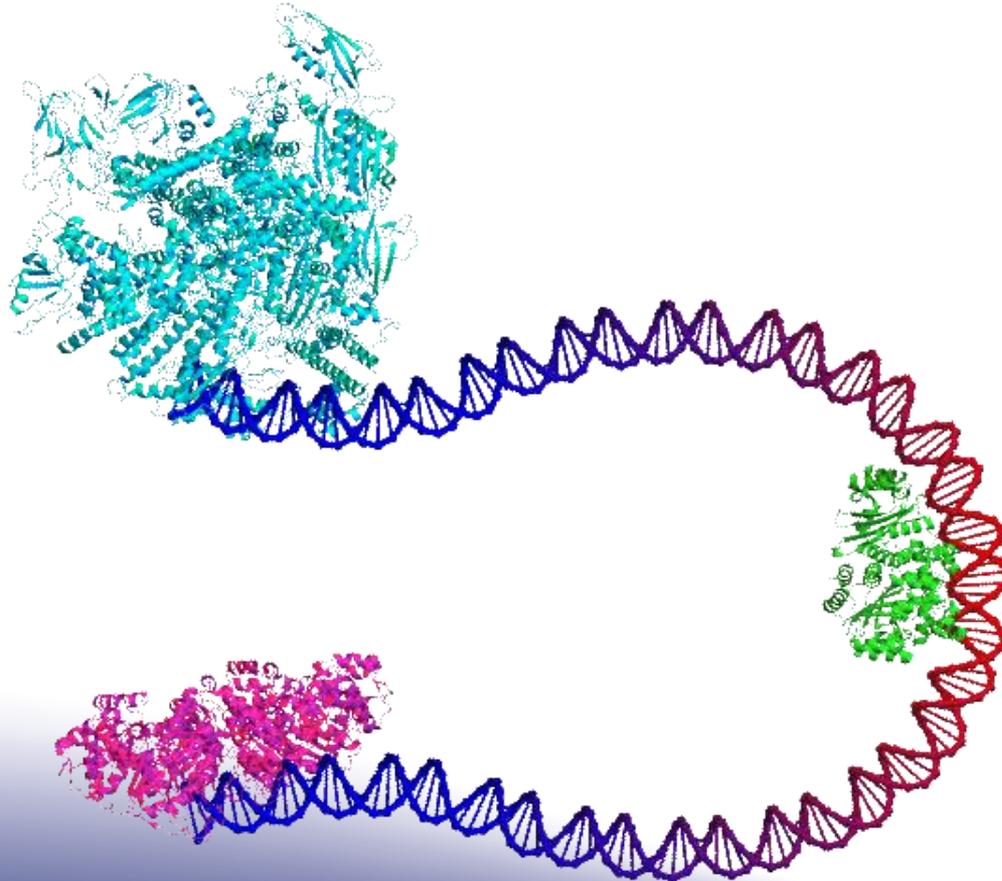
- Looping conditions mimicking bacterial  $\sigma^{54}$  promoters
- Additional model parameters:
  - $\delta \mathbf{r}$  ( $\delta \omega, \varepsilon$ )
  - $\delta \omega'$
  - $\mathbf{r}_{pol}, \mathbf{r}_{actv}$



# Excluded Volume - Reminder

- Any\* DNA binding protein can **increase** or **decrease** looping probability depending its orientation

RNA  
Polymerase

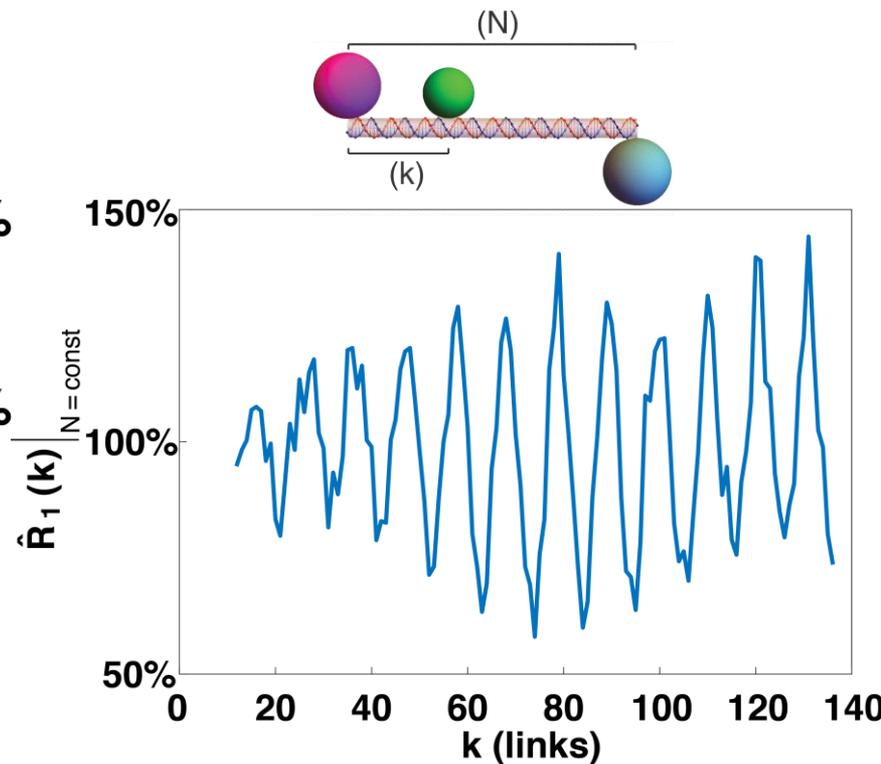
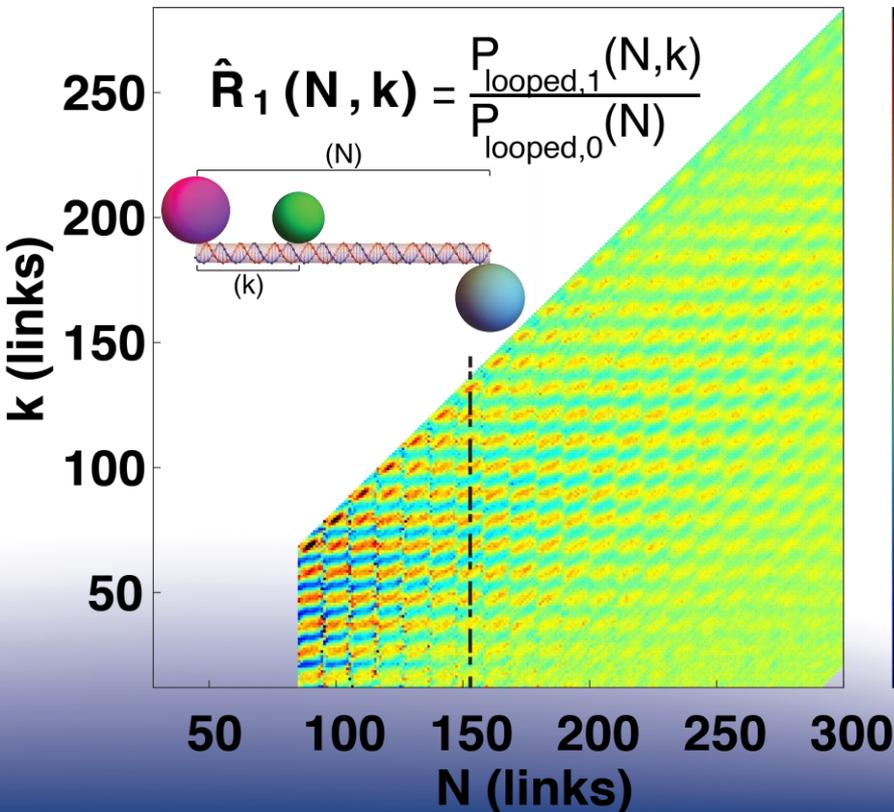


TF

Activator

# Excluded Volume

- 11 bp oscillatory pattern of up & down-regulation

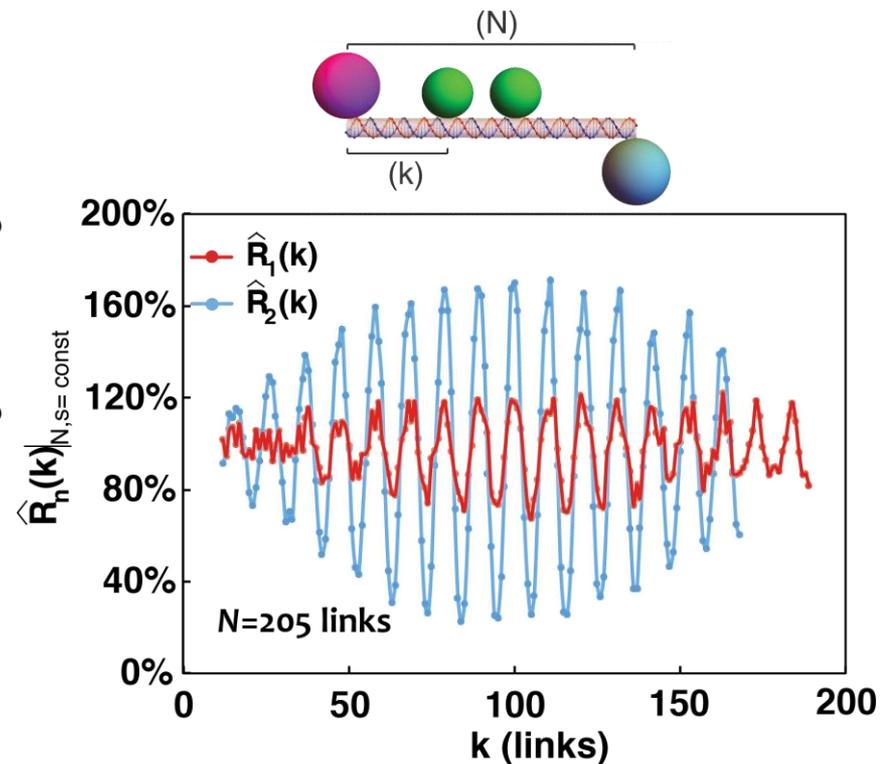
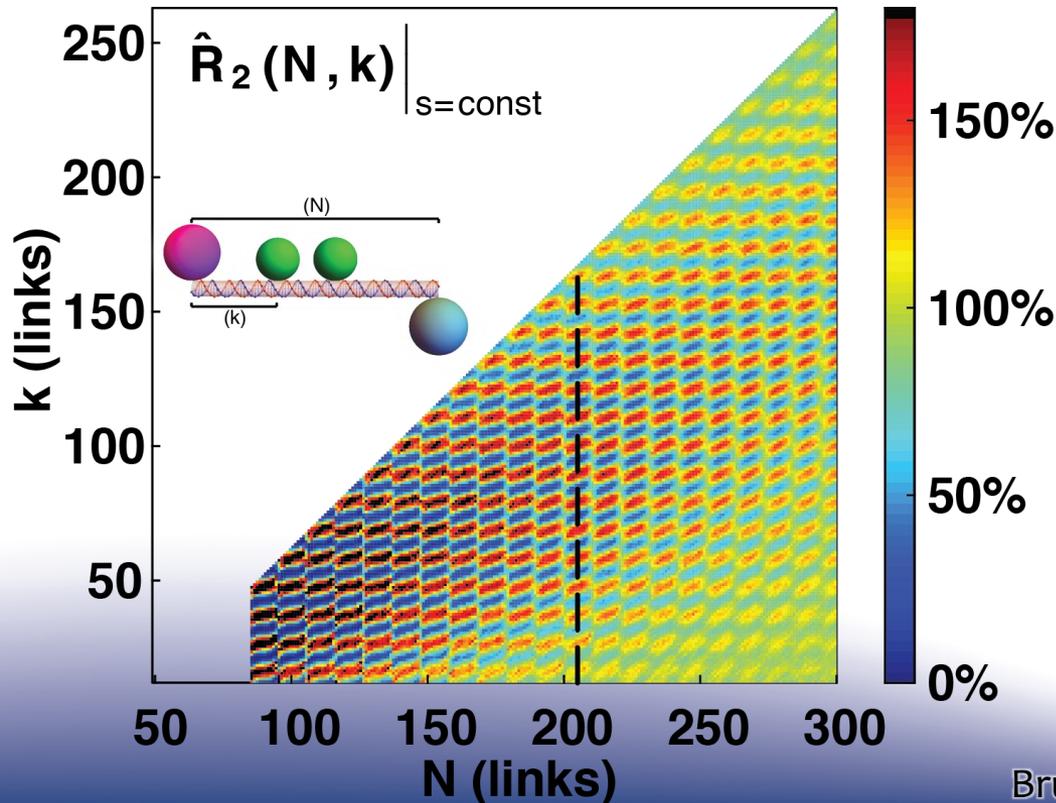


# Excluded Volume

- 11 bp oscillatory pattern of up & down-regulation

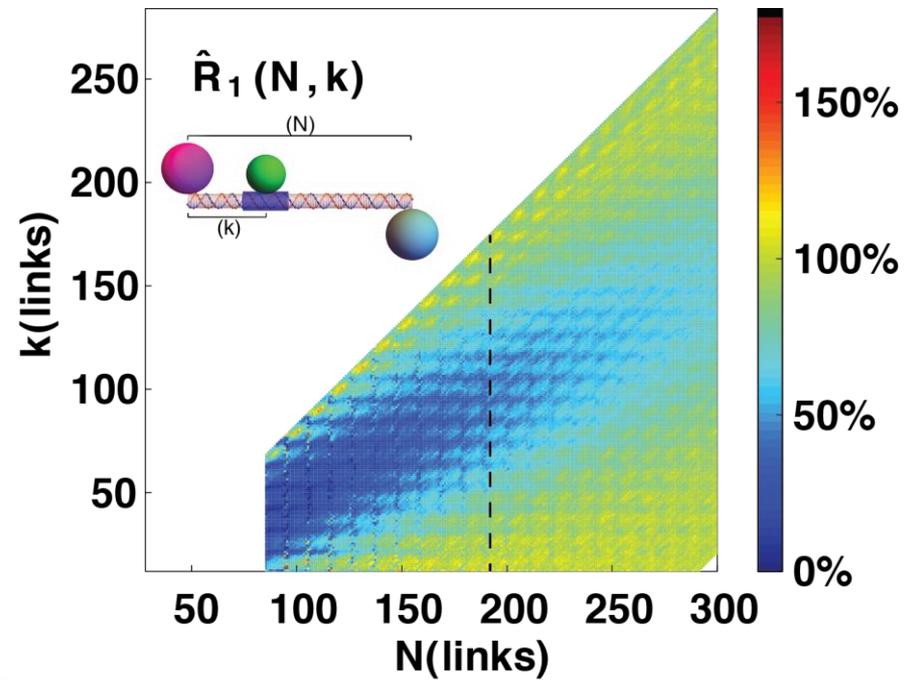
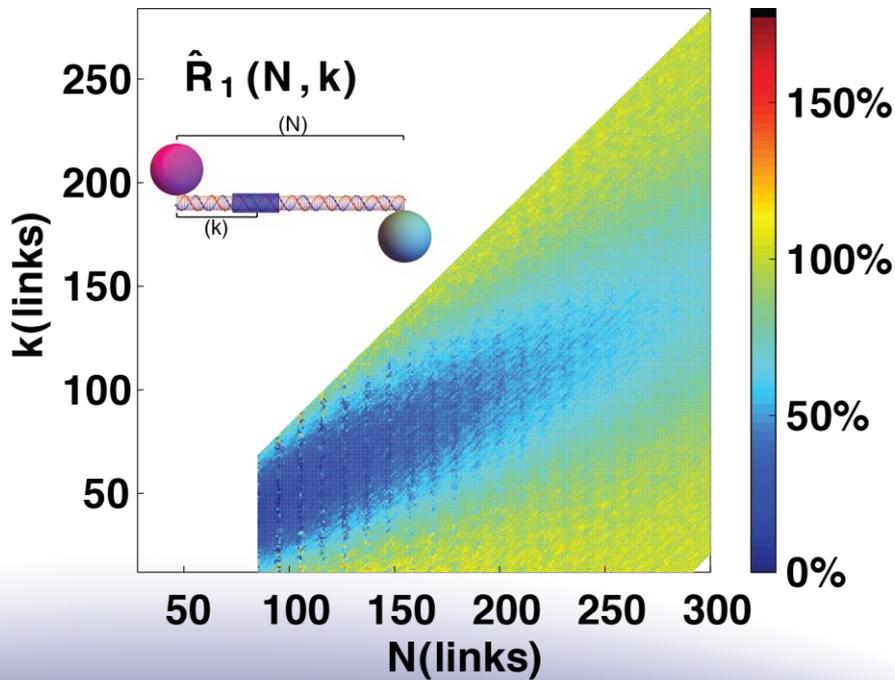


- Additive



# Stiffening

- Down-regulation, no oscillatory pattern
- Additive with the volume effect



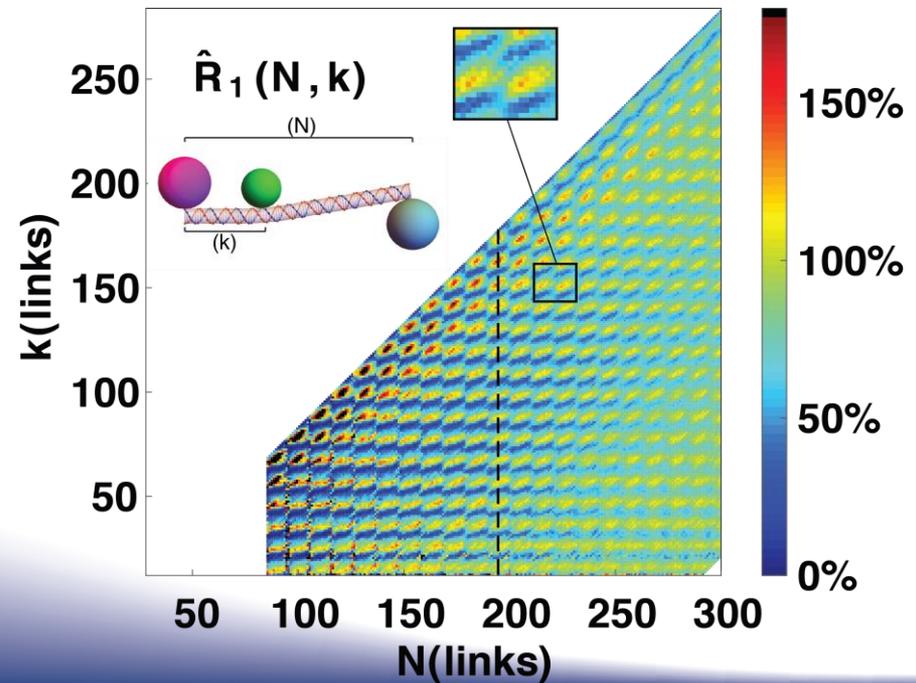
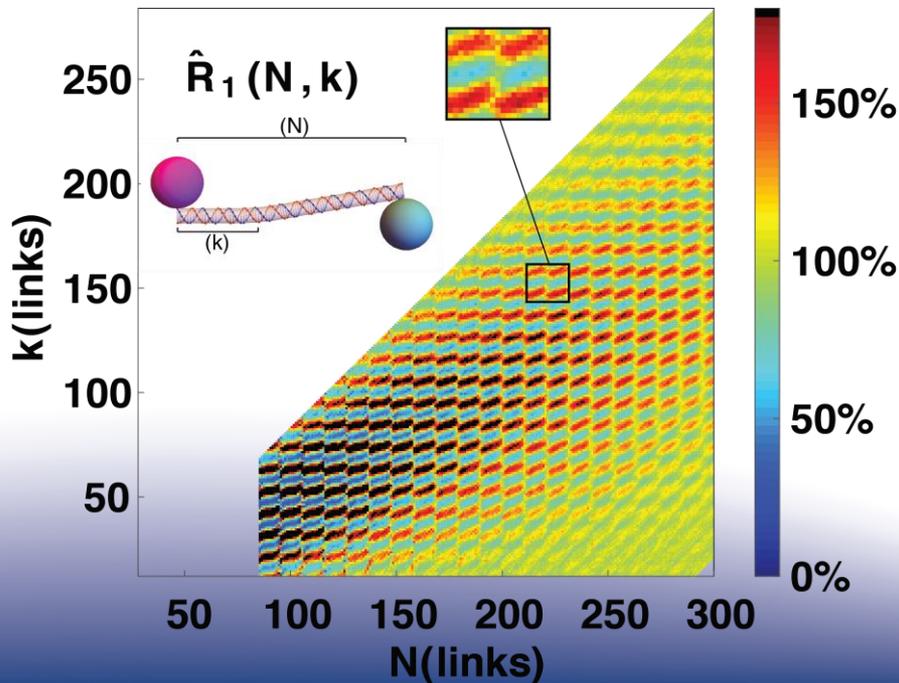
# Bending



- 11 bp oscillatory pattern of up & down-regulation



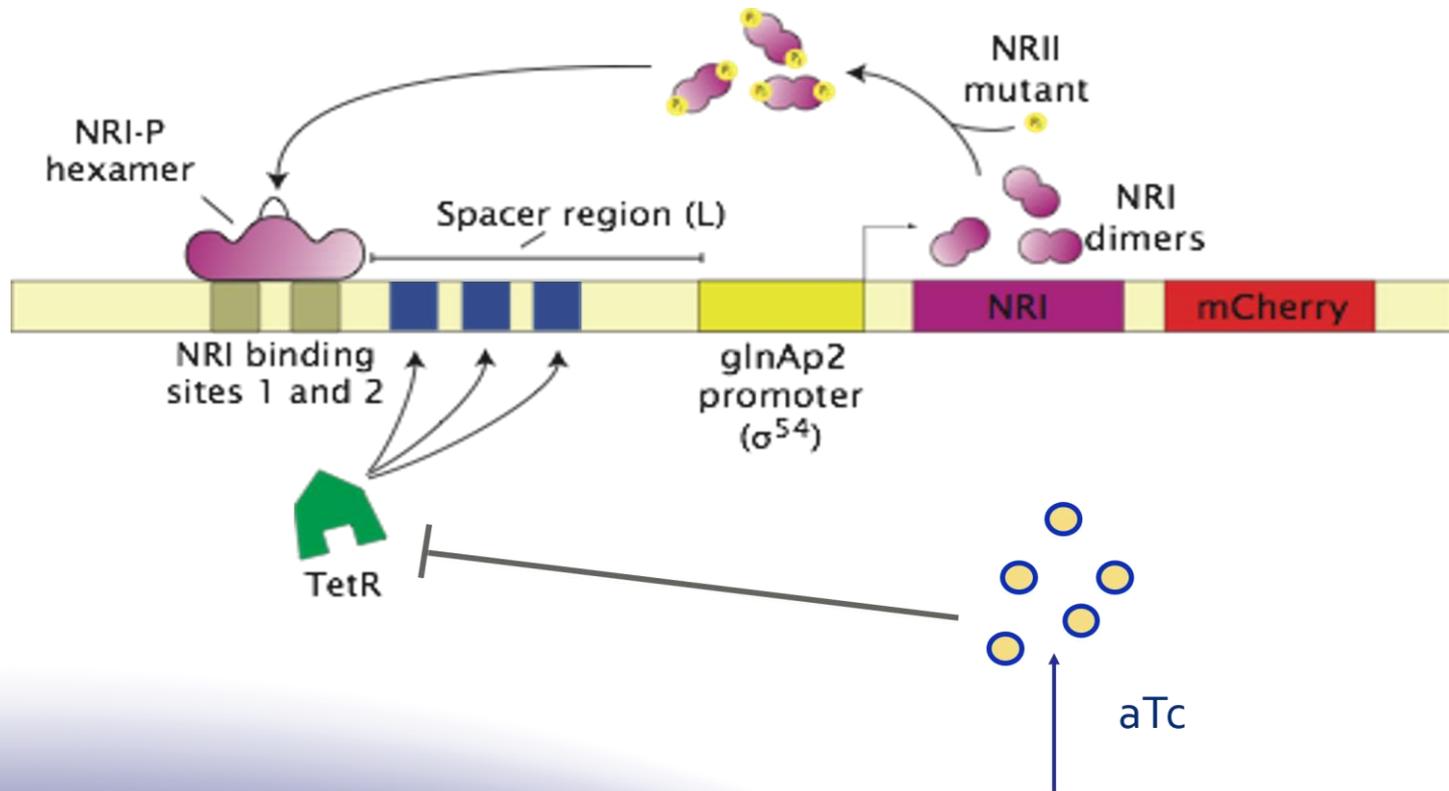
- Opposes excluded-volume



# Synthetic Biology Experiments



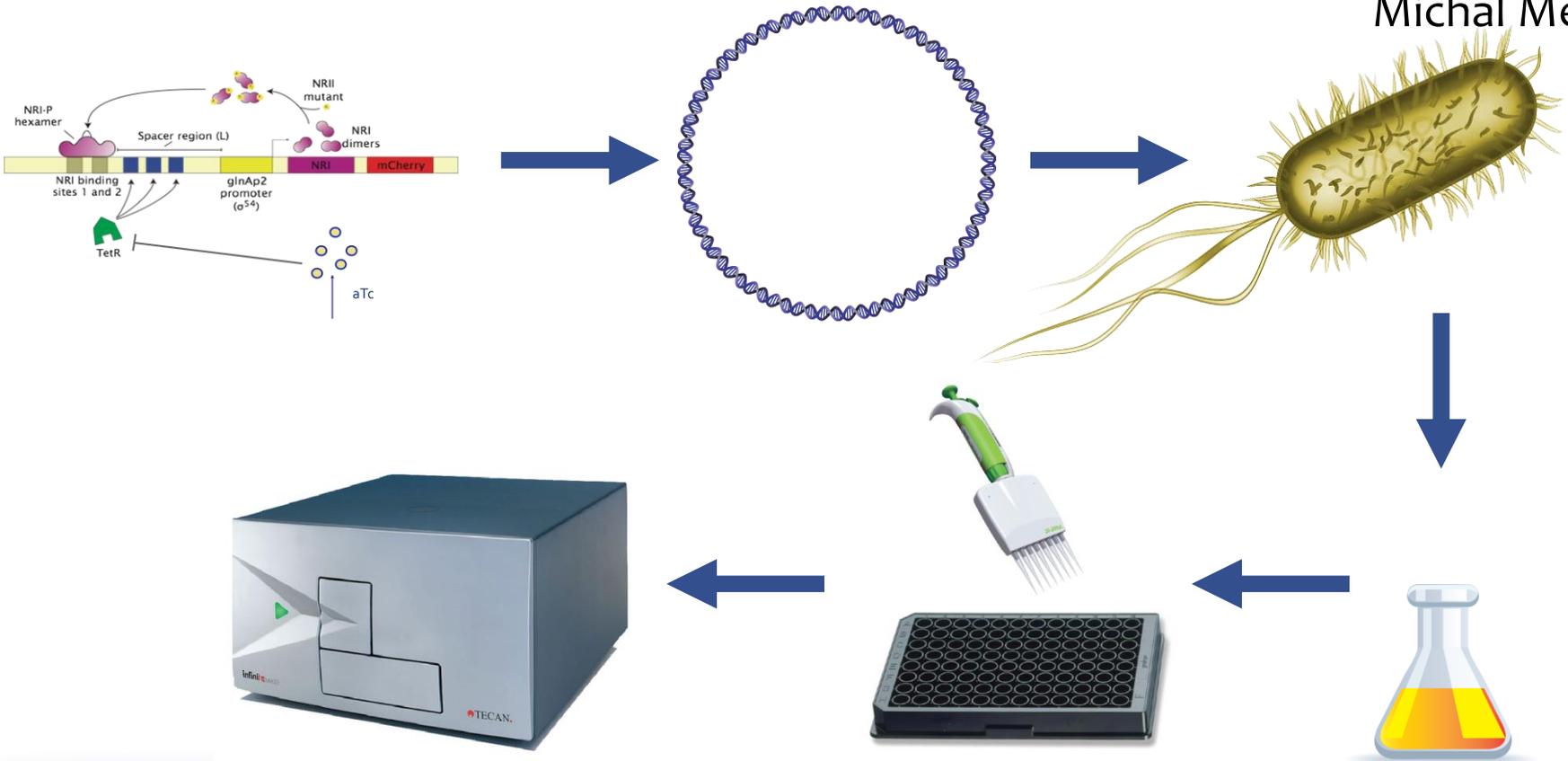
Michal Meiom



# Synthetic Biology Experiments



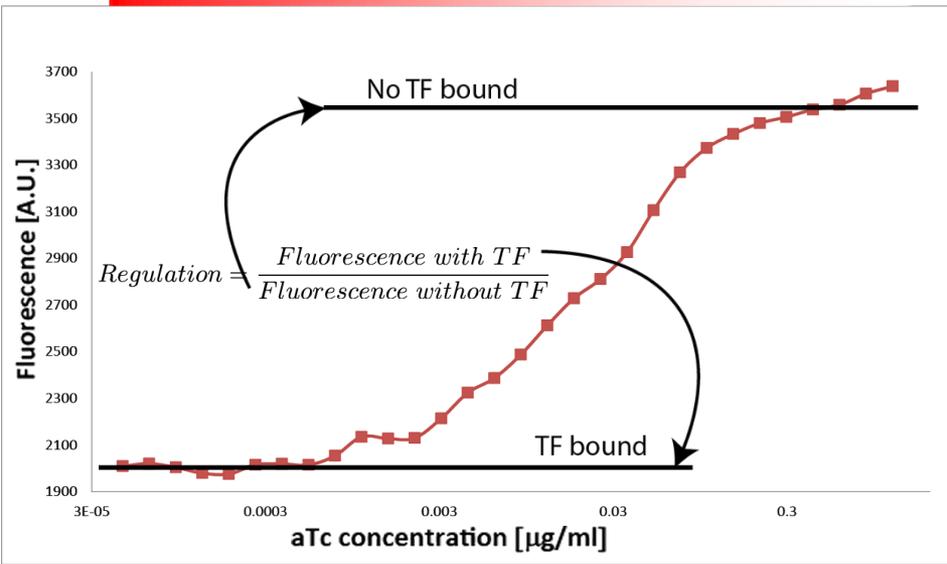
Michal Meiom



# Measurements

- OD normalized fluorescence
- Fluorescence ratio  $\approx$  looping probability ratio

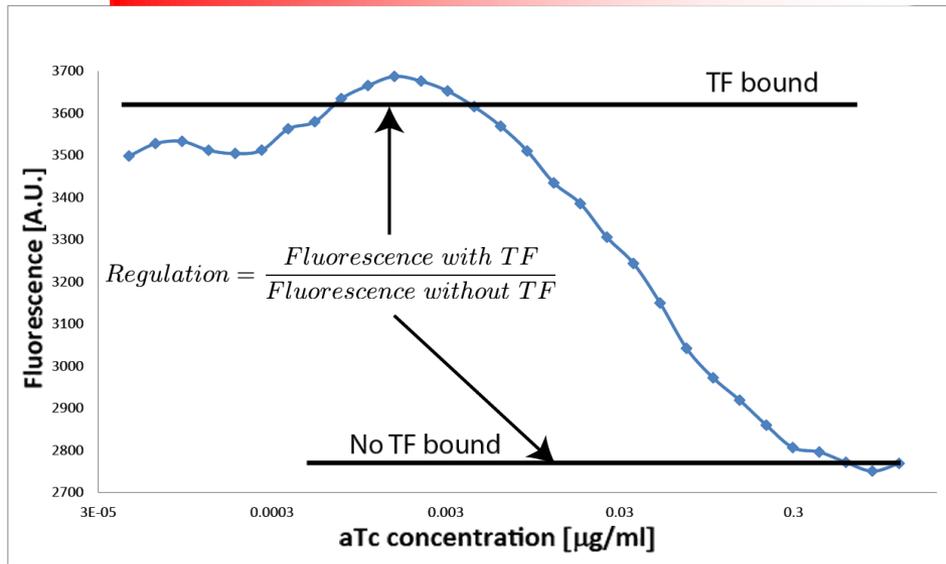
TetR



TetR, k=64 bp

Down-regulation

TetR



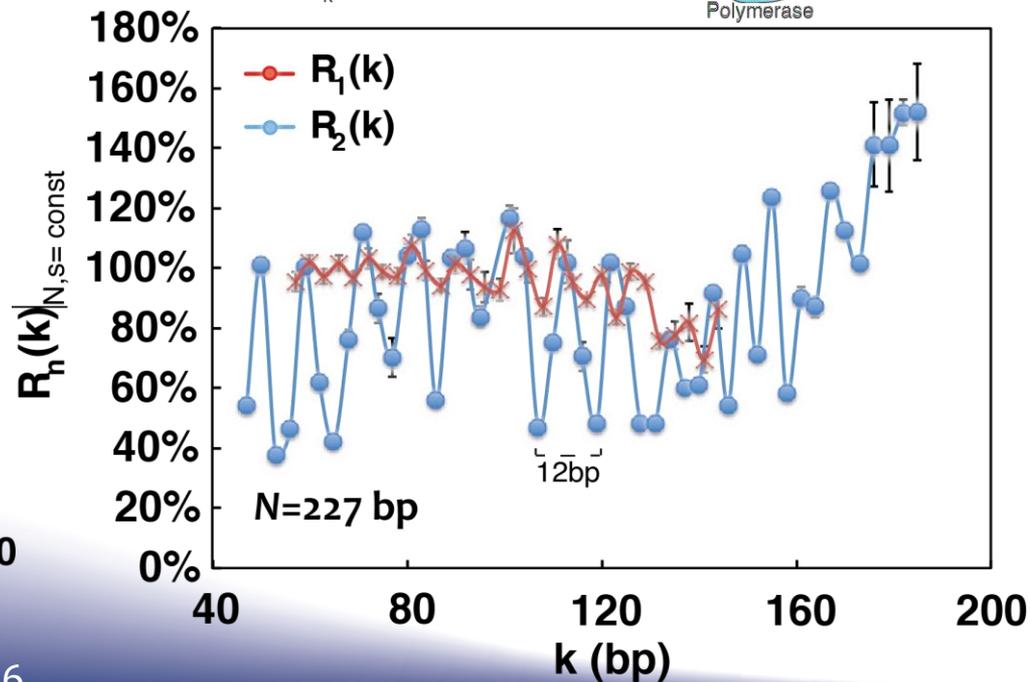
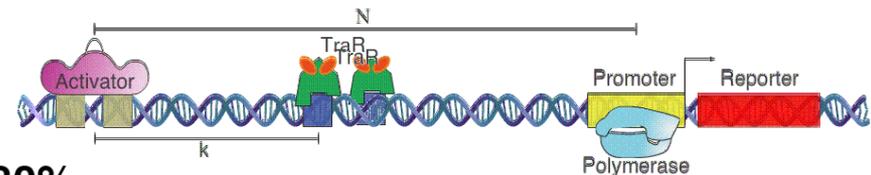
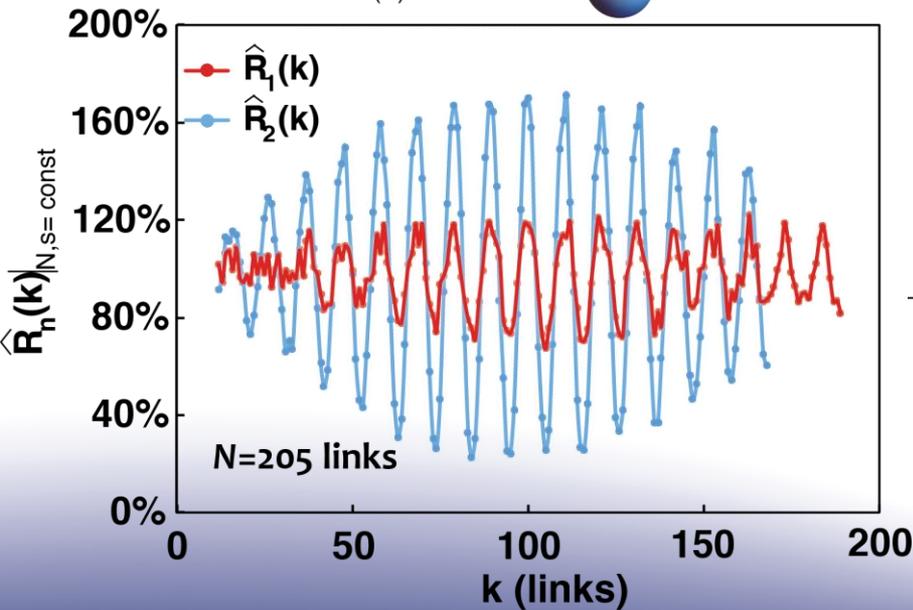
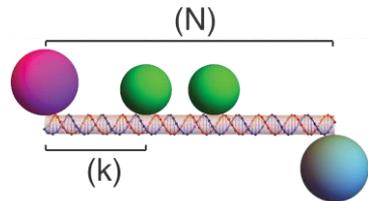
$\Delta k=25$ bp

TetR, k=89 bp

Up-regulation

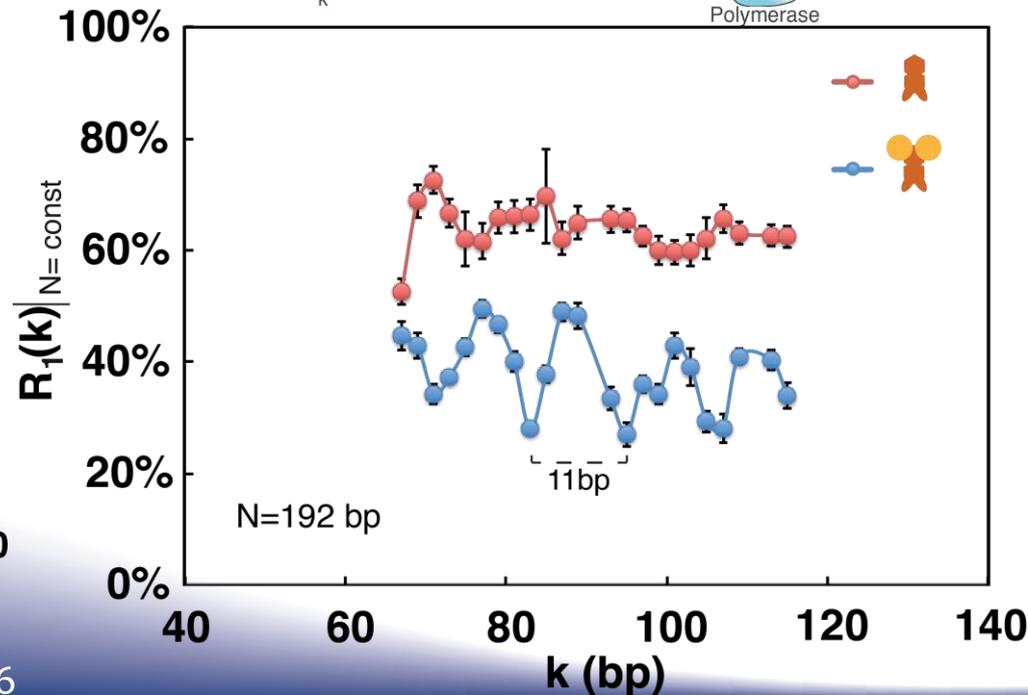
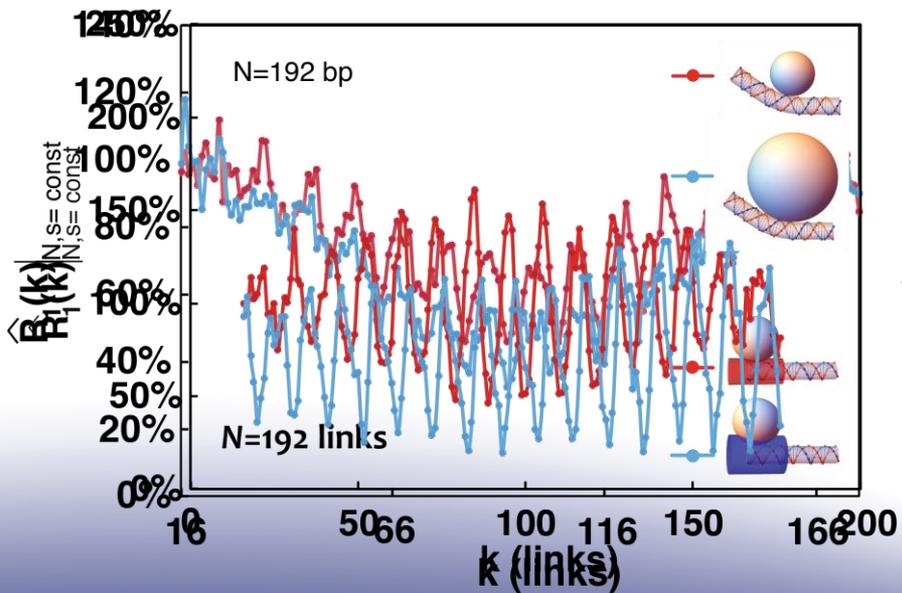
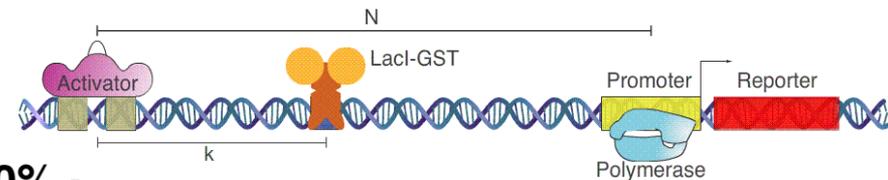
# TraR Experimental Results

- 11 bp oscillatory pattern of up & down-regulation
- Additive effect



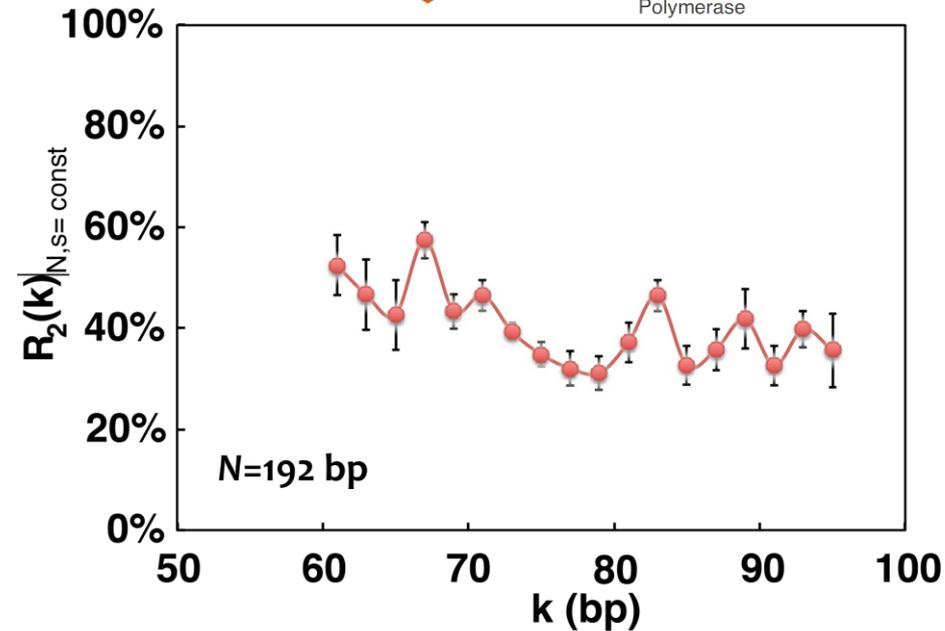
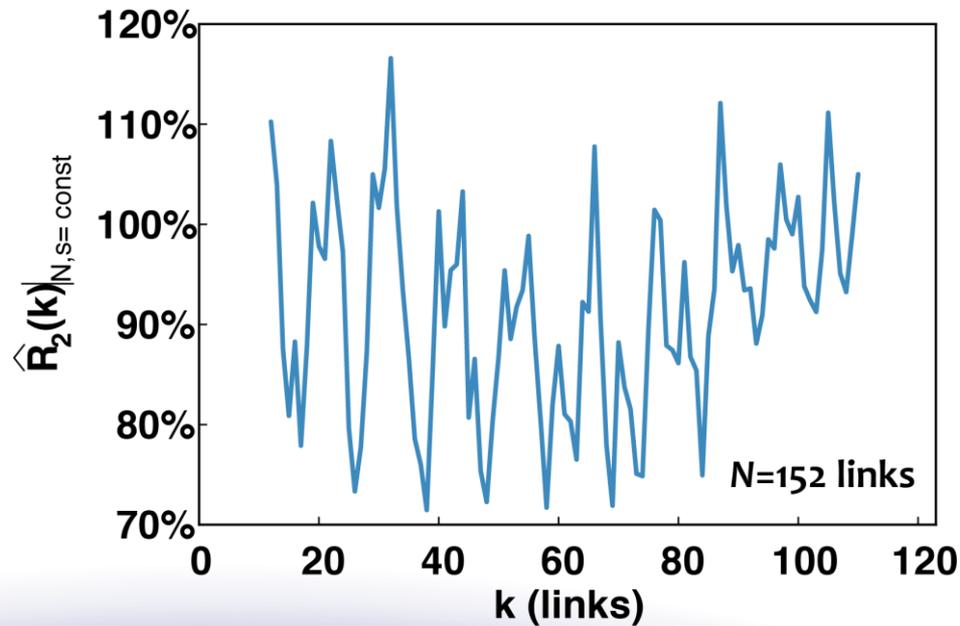
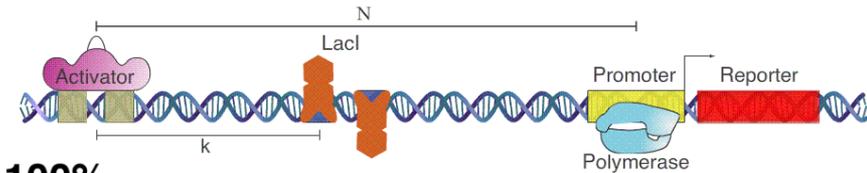
# LacI Experimental Results

- Larger volume  $\Rightarrow$  larger amplitude
- Oscillations phase flip
- Shift towards down regulation

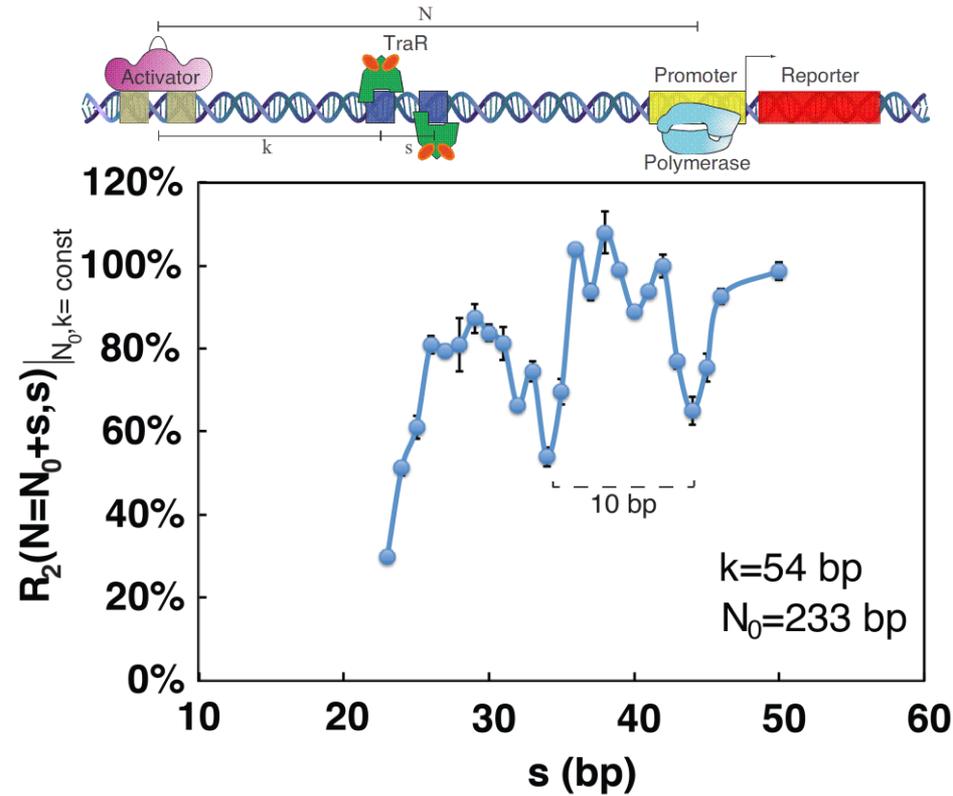
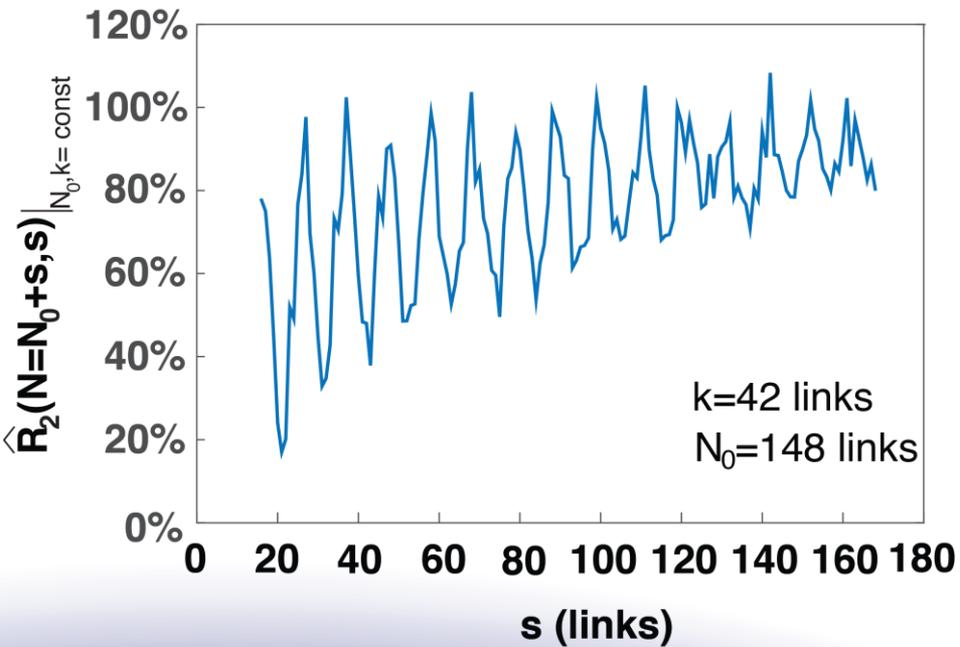


# Multiple TFs – Out of Phase

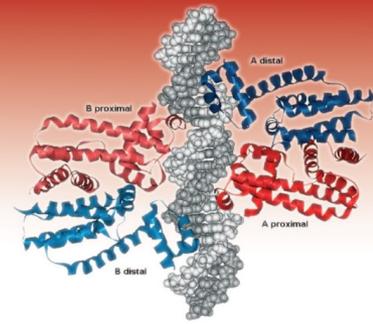
- Deleterious, weak down-regulation with  $\frac{1}{2}$  DNA helical repeat periodicity.



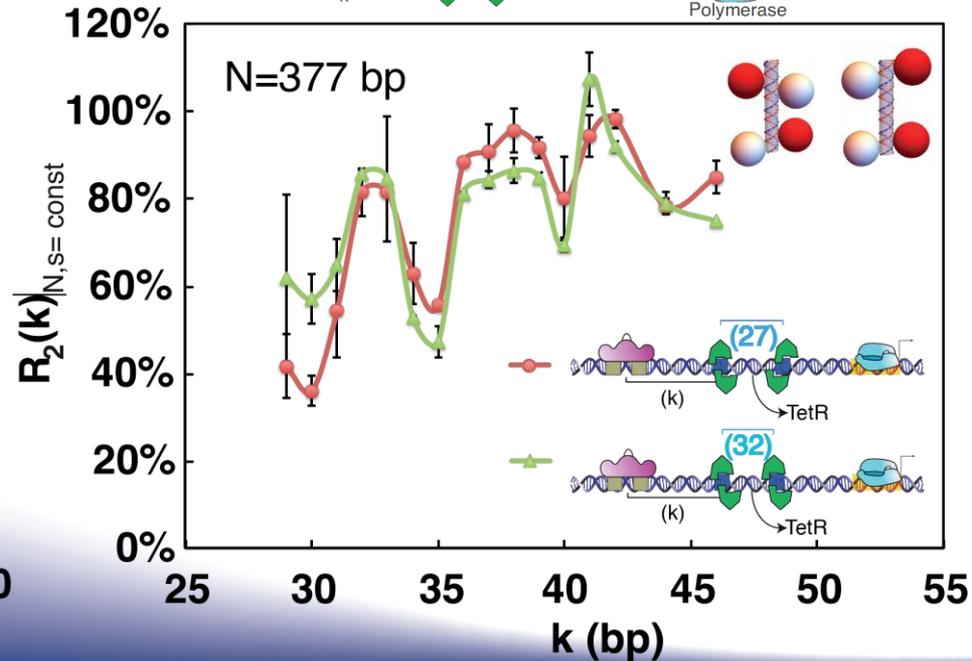
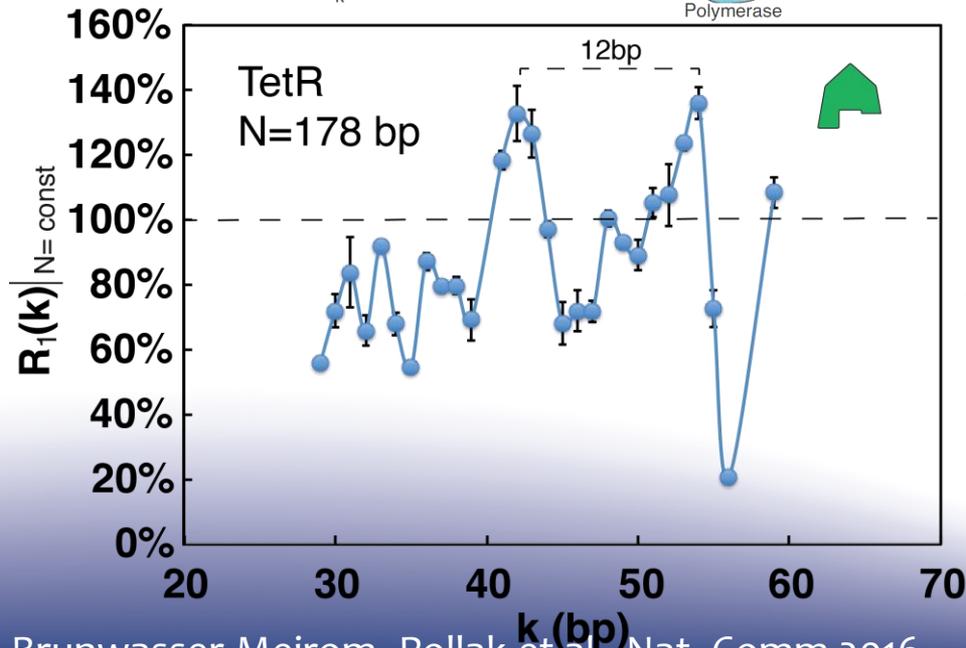
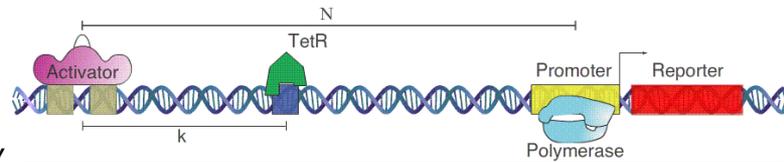
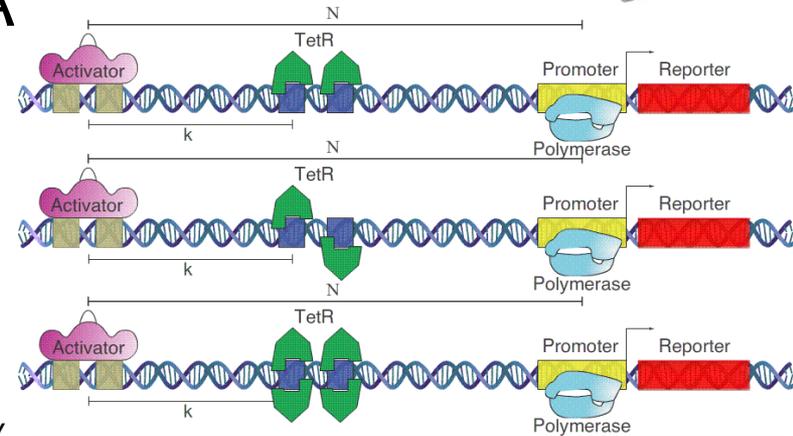
# Multiple TFs



# Structural Insights



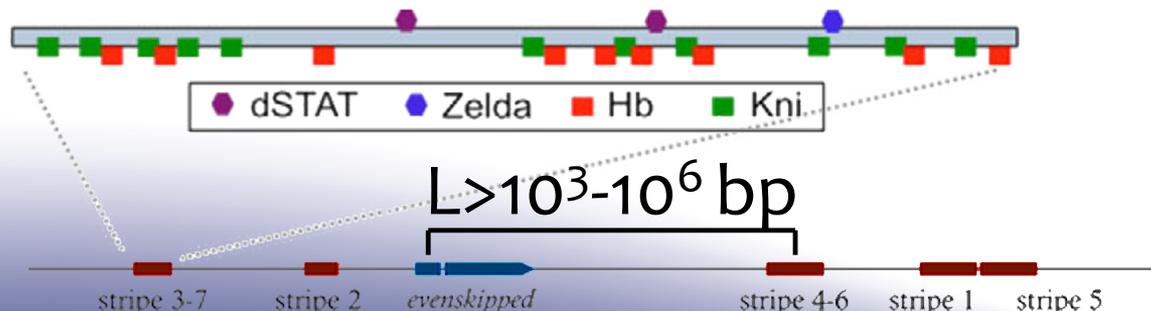
- Results suggest TetR binds DNA similar to its homolog QacR.



# Long-Range Looping Results

# Short vs. Long range

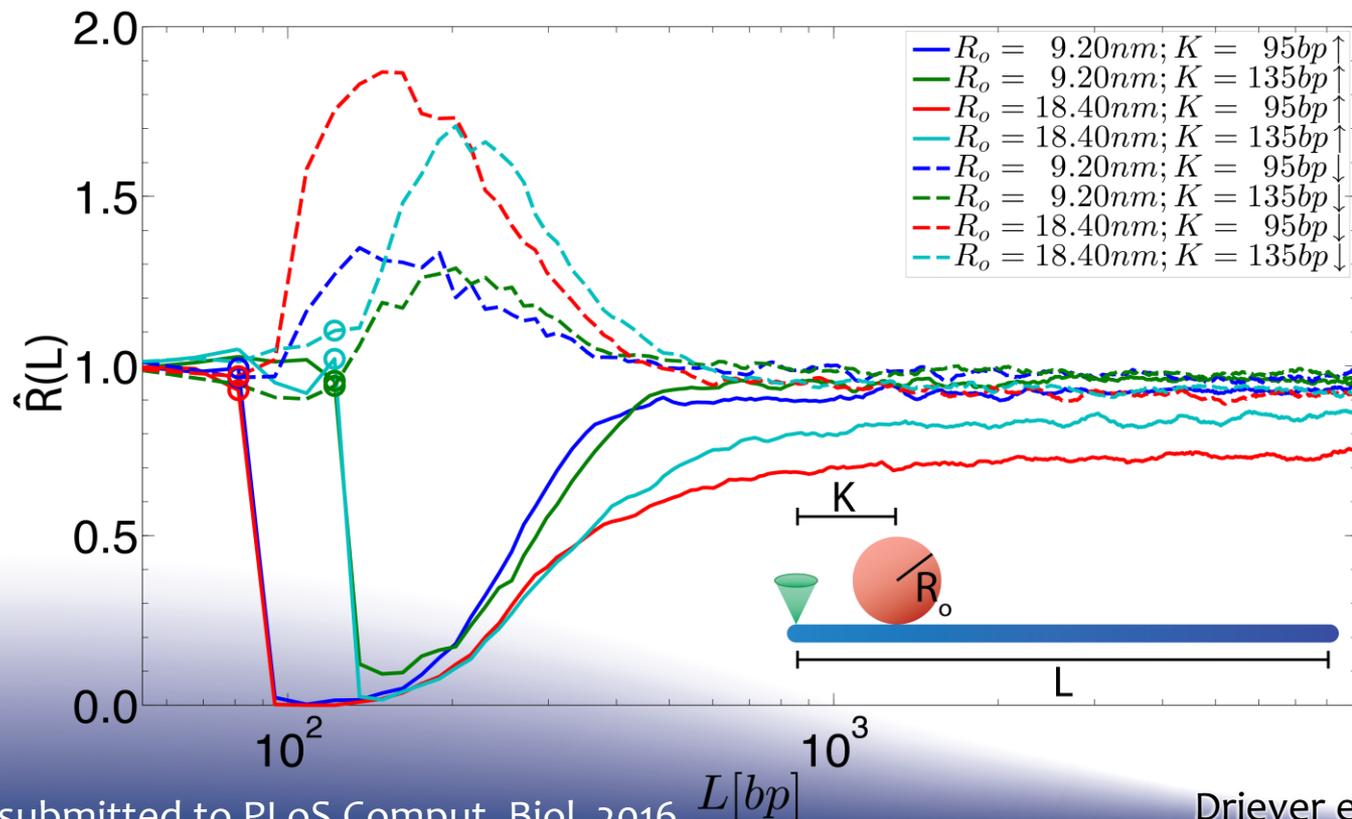
- Short range (elastic regime) ( $\sim 10^2$ bp)
  - Bending
  - Stiffening
  - Excluded volume
- Long range (entropic regime) ( $> 10^3$ bp)
  - ~~Bending~~
  - ~~Stiffening~~
  - Excluded volume





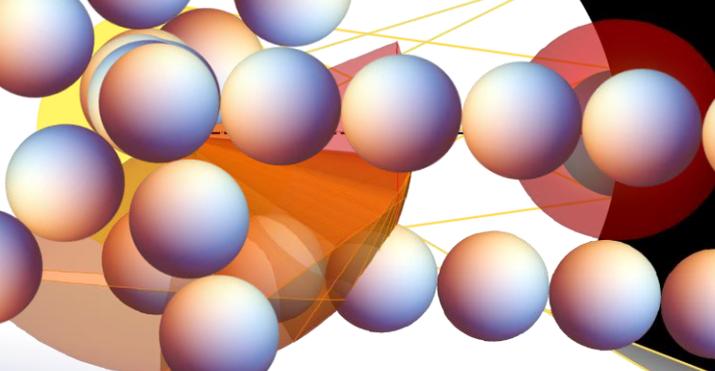
# Excluded Volume Effect

- The effect is relatively constant at long range
- Always down-regulation
- Only sizable for TFs in-phase with the looping volume



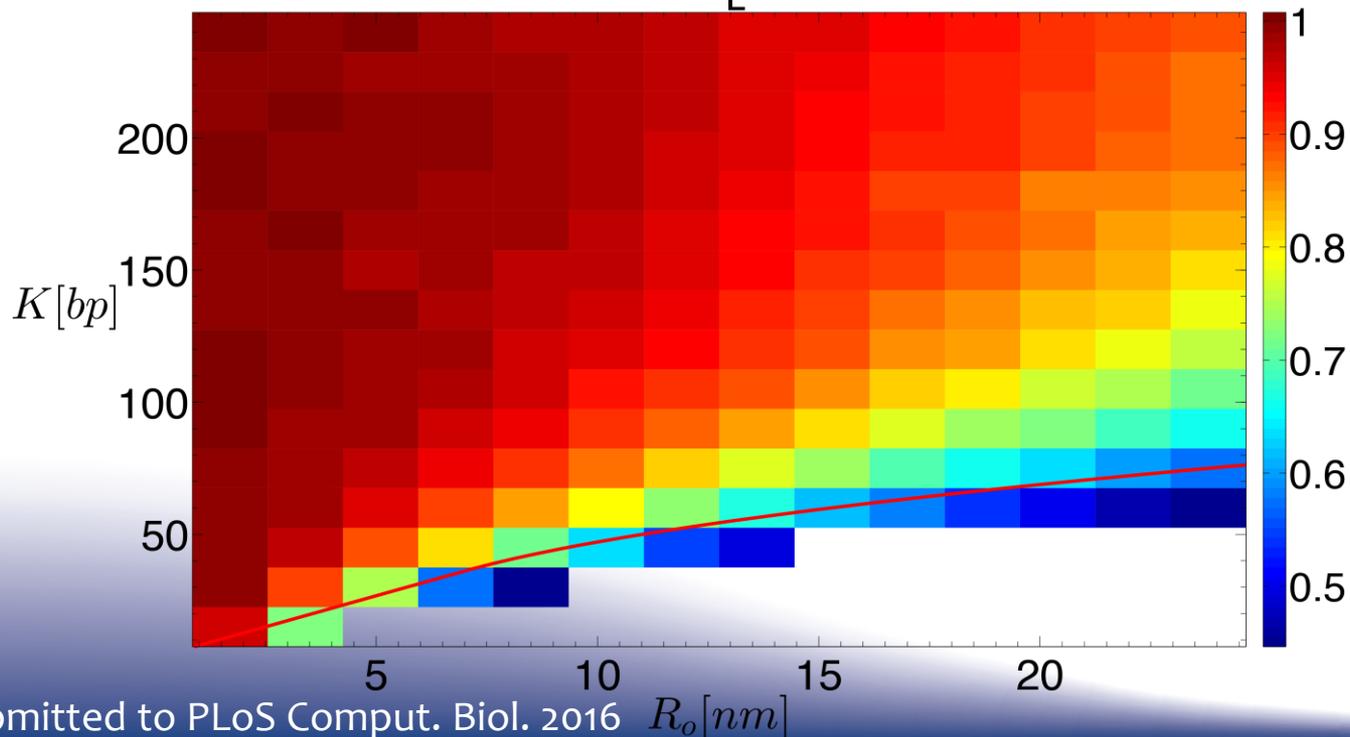
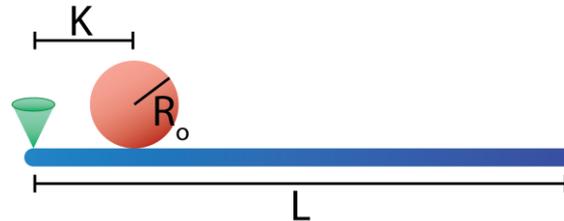
# Mechanism - Eclipsing

- The  $\text{C-H}$  bonds are aligned so that they are seen from all directions uniformly
- The eclipsing angle  $\theta$  is a portion of the light depending on the distance from the lens and the solid angle at the viewing volume



# Eclipsing Effect

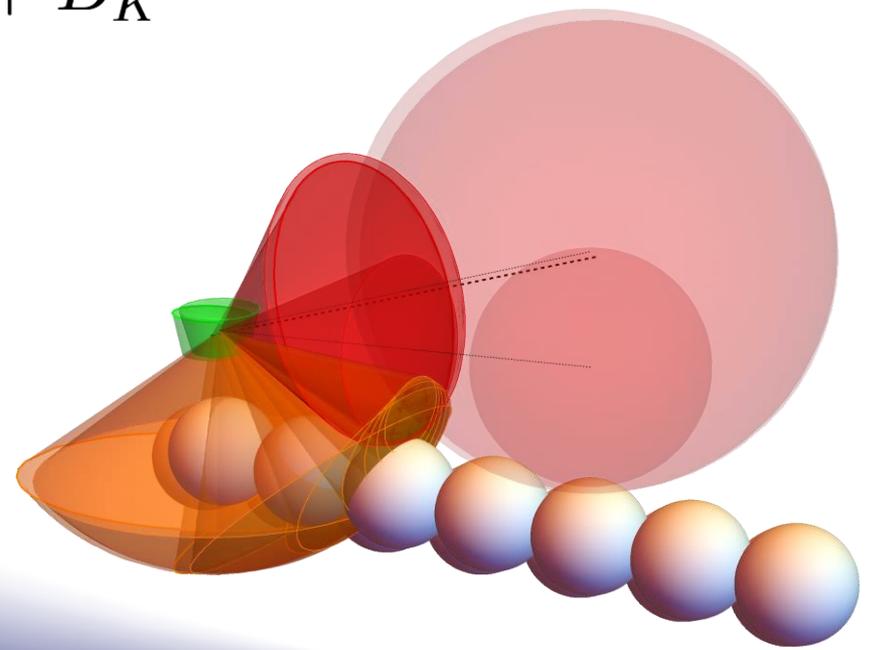
- The effect is stronger with larger TF
- The effect dies out fast as the protrusion is moved away from the looping volume



# Mechanism contd.

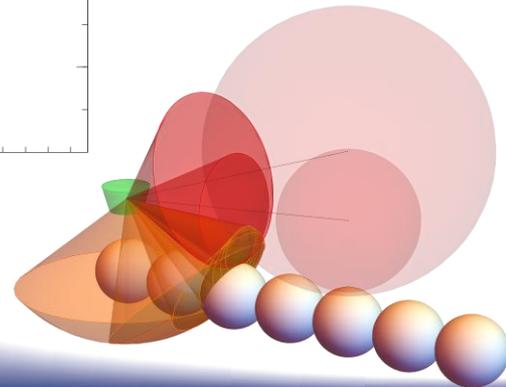
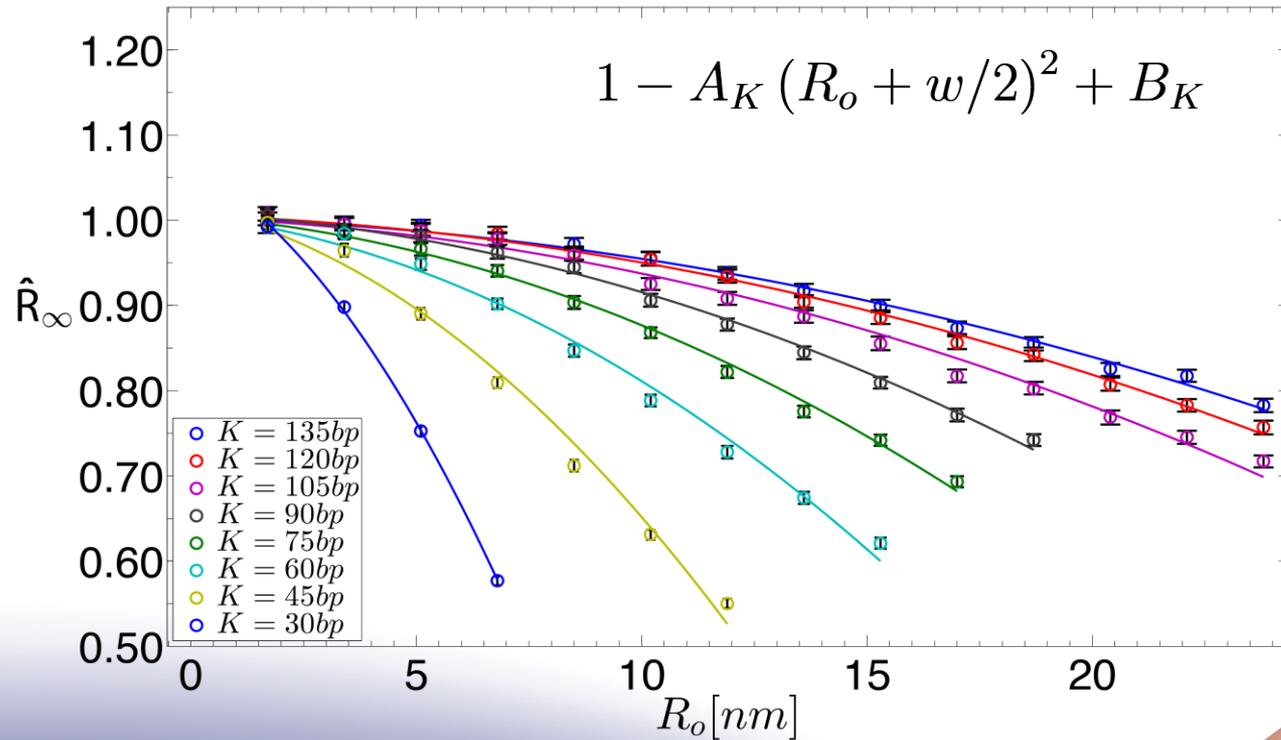
- Have to take the chain solid angle into account
- Distance is not straightforward
- $\hat{R}$  can be approximated for TF constant location

$$\hat{R} \approx 1 - A_K (R_o + w/2)^2 + B_K$$



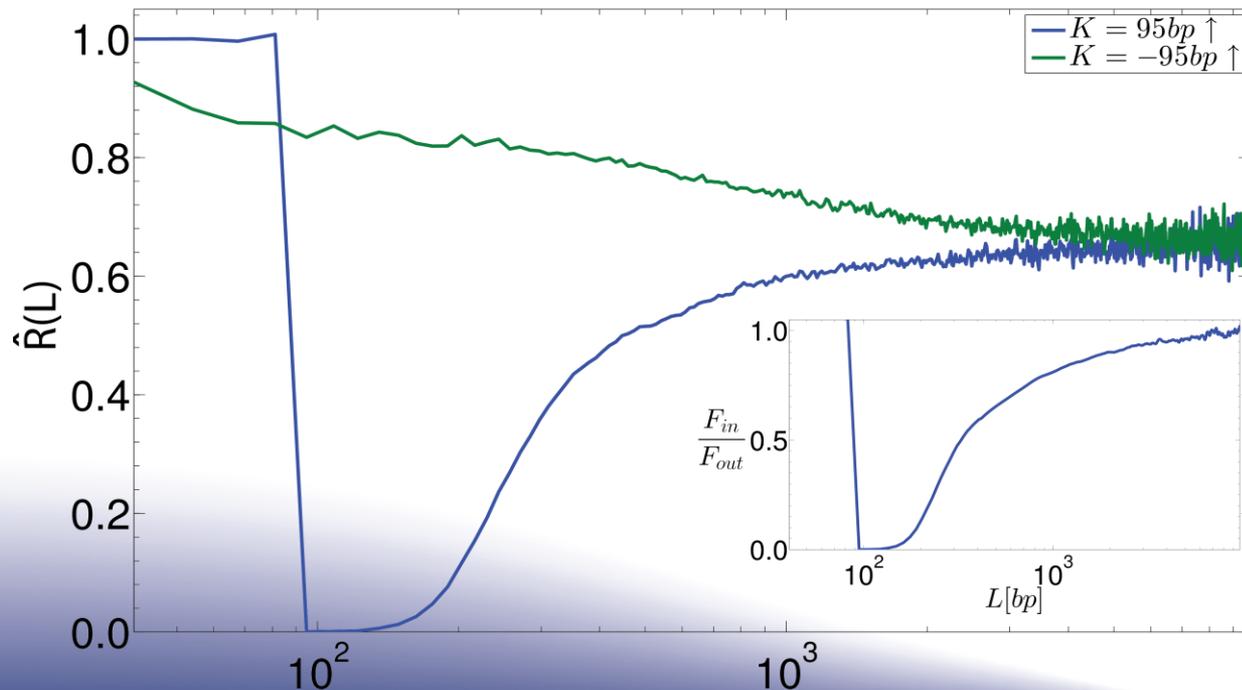
# Mechanism contd.

- $R^2 > 0.99$  for fit to quadratic functions in  $R_o$ .

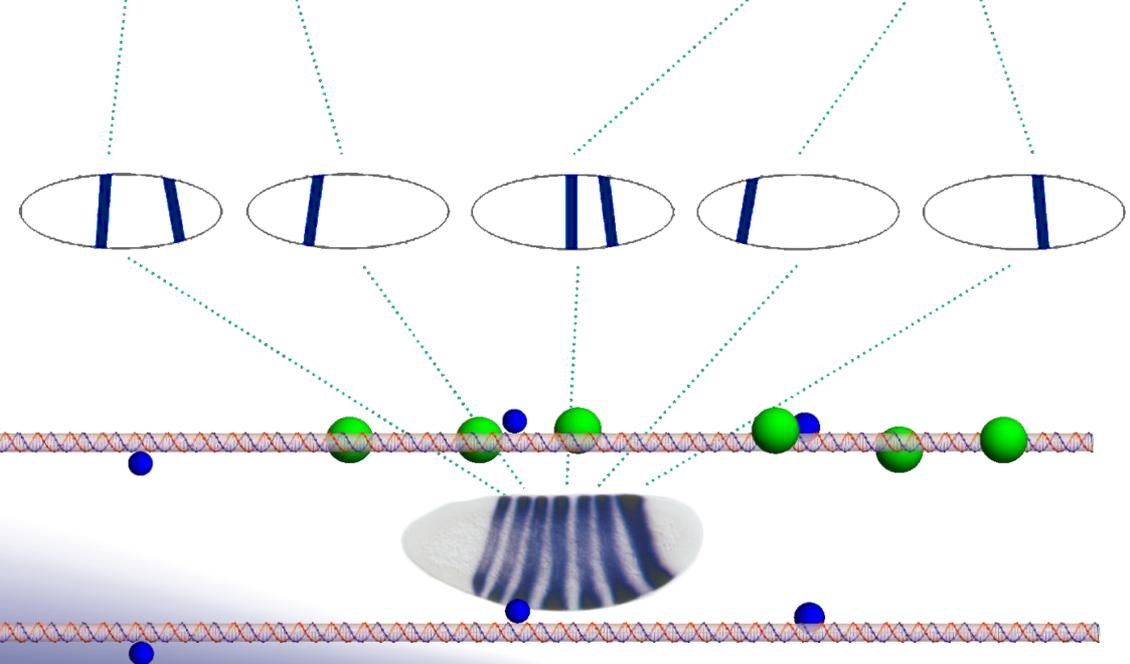
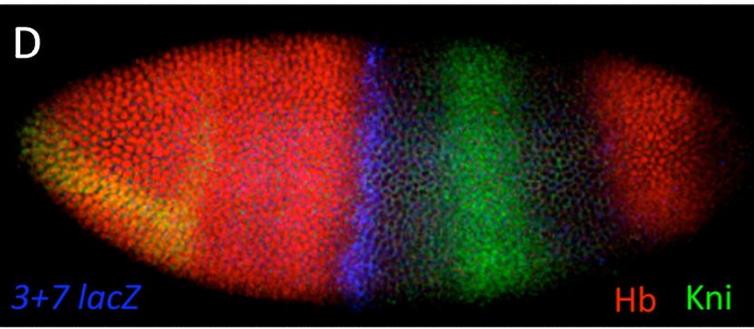
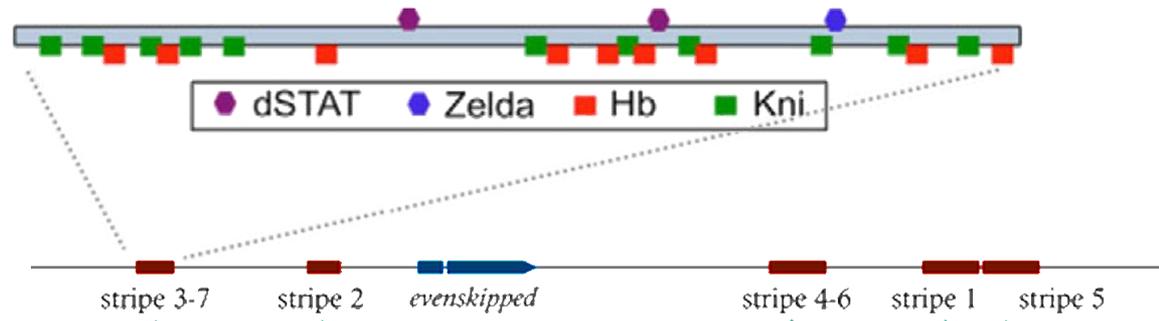
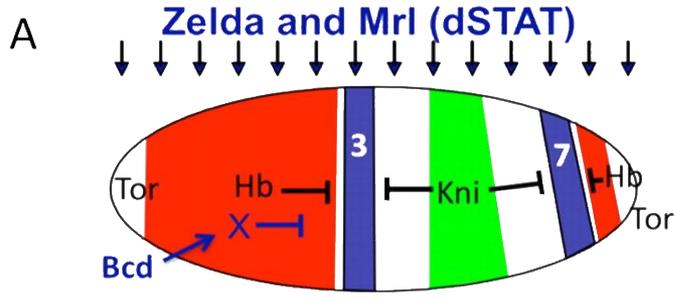


# Upstream / Downstream

- The effect is independent of whether the TF is located upstream or downstream from the driver



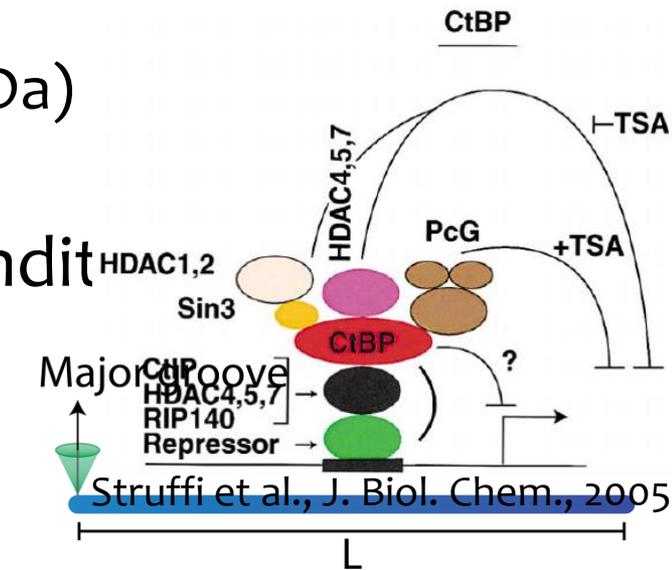
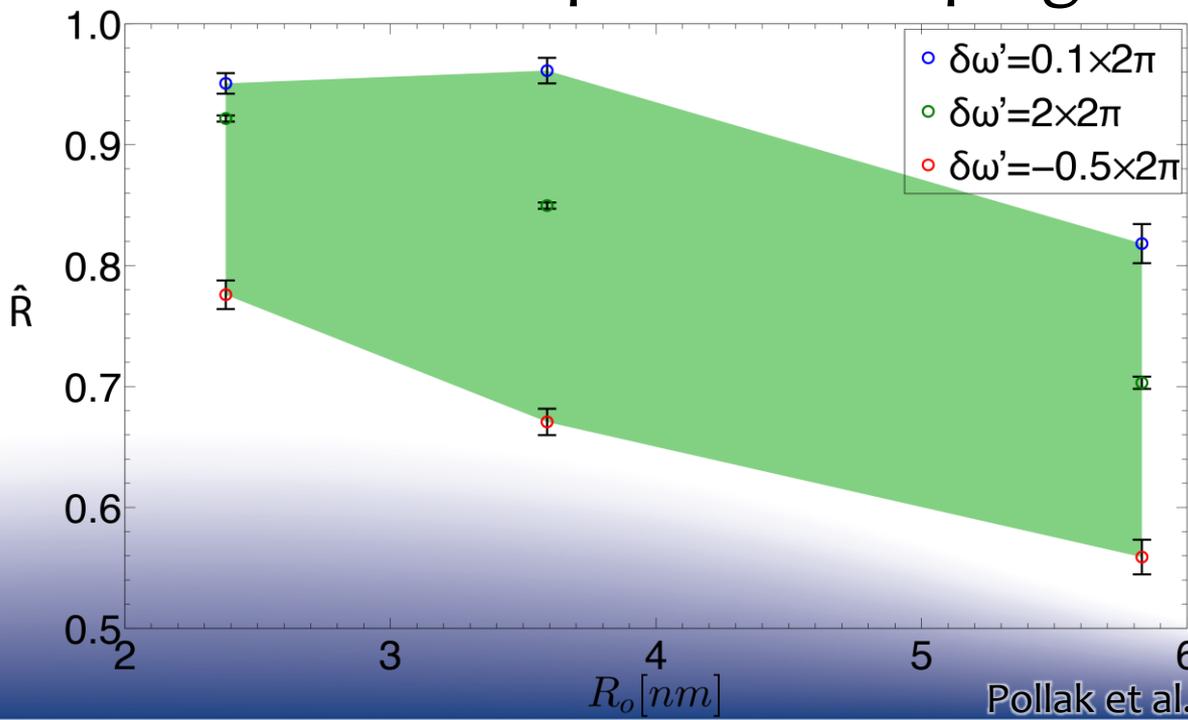
# eve 3/7



- State 1:
- State 2:

# eve 3/7 contd.

- Tested three possible repressor complex sizes:
  - Knirps alone (46 kDa)
  - Knirps bound to CtBP dimers (130 kDa)
  - Full putative 450 kDa complex
- Tested three possible looping conditions



# Conclusions

- Self-avoiding wormlike chain model can explain looping-based regulation
- Excluded volume can generate long-range repression
- Model predicts reduction in probability of looping for Knirps fully occupied eve 3/7 enhancer.



# Thanks



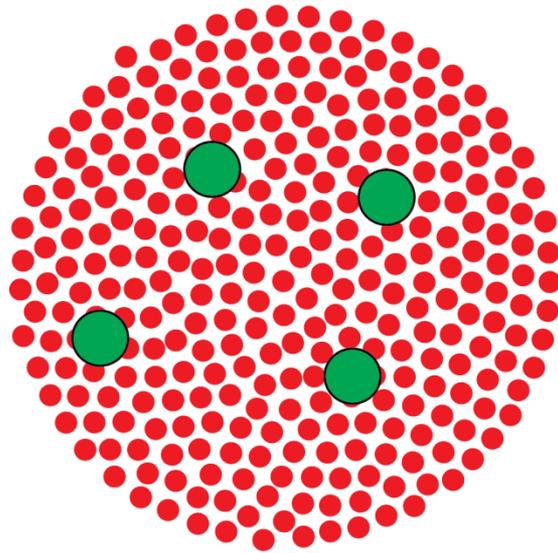
- Roei Amit
- Michal Meirom
- Sarah Goldberg
- Lior Levy
- Orna Atar
- Everybody else
- My Wife
- Our Parents
  
- RBNI – Daniel
- RBNI



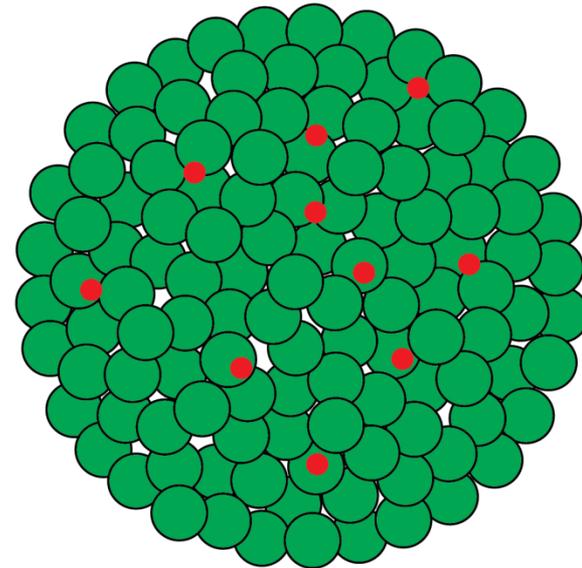


# Uniform Sampling

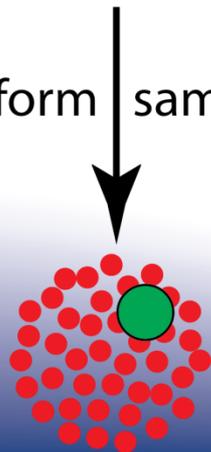
Geomerty



Reality



Uniform sampling

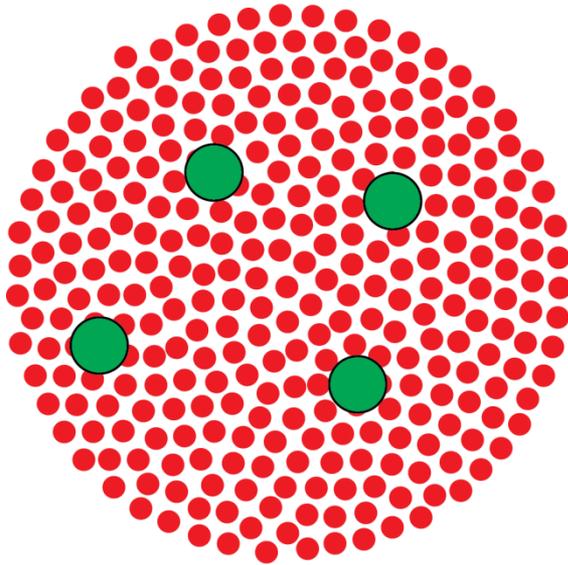


$$P_{chain} \propto e^{-\beta E_{chain}}$$

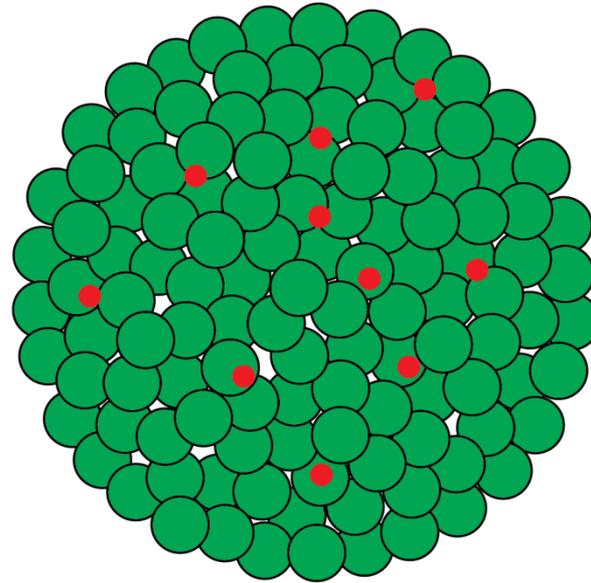


# Importance Sampling

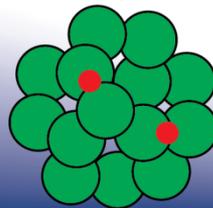
Geomerty



Reality



Smart sampling



# Sampling Bias

Would like to know:  $\langle v \rangle = \int v(\mathbf{x}) \pi(\mathbf{x}) d\mathbf{x} = \frac{\int v(\mathbf{x}) \exp(-\beta E(\mathbf{x})) d\mathbf{x}}{Z}$

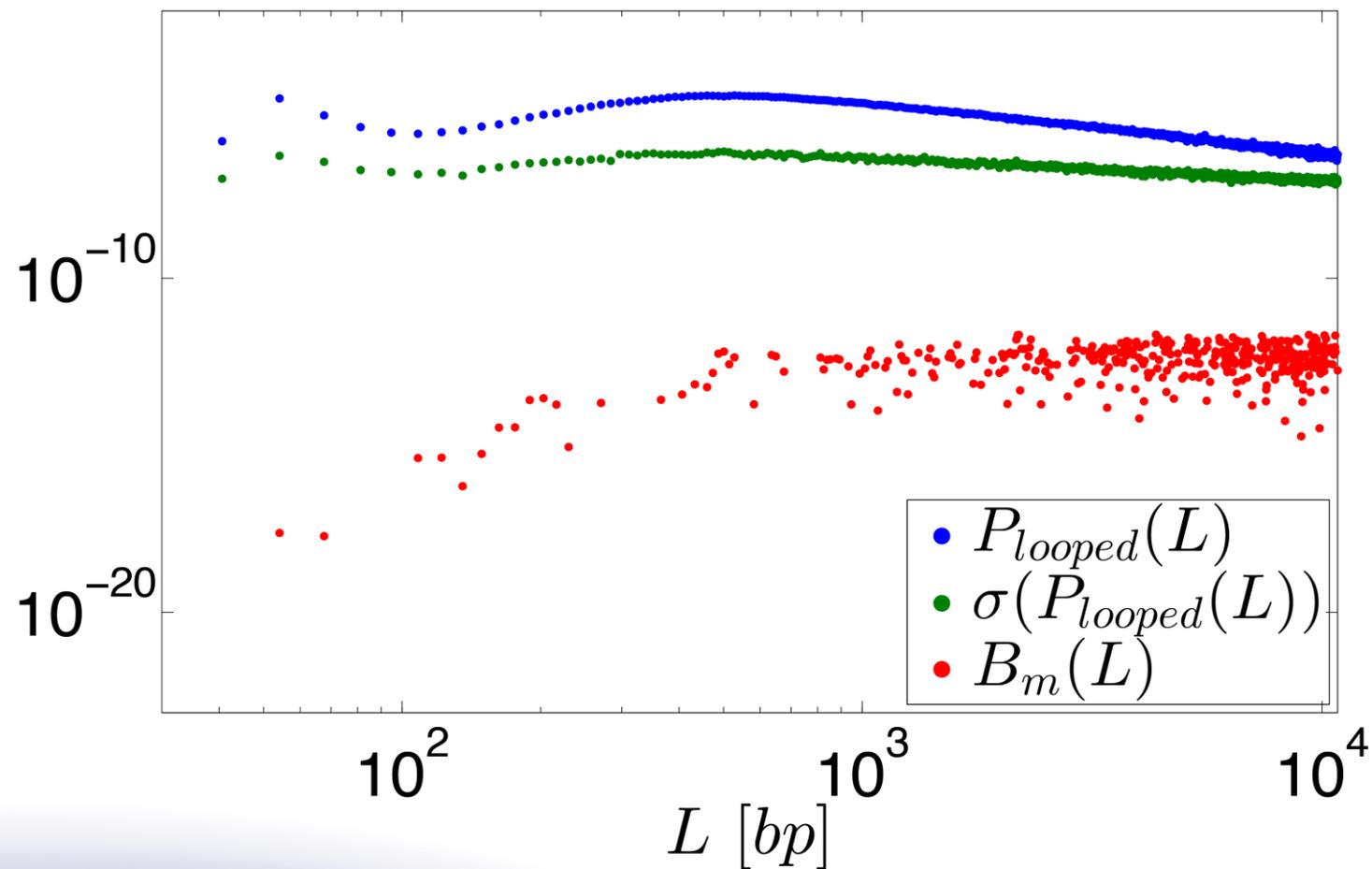
Estimate from ensemble:  $\langle \hat{v} \rangle = \frac{\sum_{i=1}^m w(\mathbf{x}^{(i)}) v(\mathbf{x}^{(i)})}{\sum_{i=1}^m w(\mathbf{x}^{(i)})}$

Should have used:  $\langle \tilde{v} \rangle = \frac{1}{Z} \frac{1}{m} \sum_{i=1}^m w(\mathbf{x}^{(i)}) v(\mathbf{x}^{(i)})$

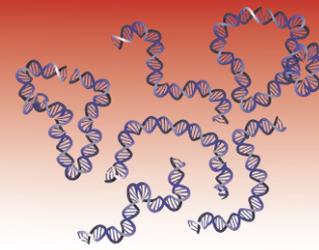
Sampling bias:  $B_m = E[\langle \hat{v} \rangle] - \langle v \rangle = -\frac{1}{mZ^2} \left( \int (v(\mathbf{x}) - \langle v \rangle) w(\mathbf{x}) \exp(-\beta E(\mathbf{x})) d\mathbf{x} \right)$

Practical bias estimator:  $B_m = E[\langle \hat{v} \rangle] - \langle v \rangle = -\frac{\text{covar}(\langle \hat{v} \rangle \hat{Z})}{Z}$

# Sampling Bias



# Monte-Carlo Simulations



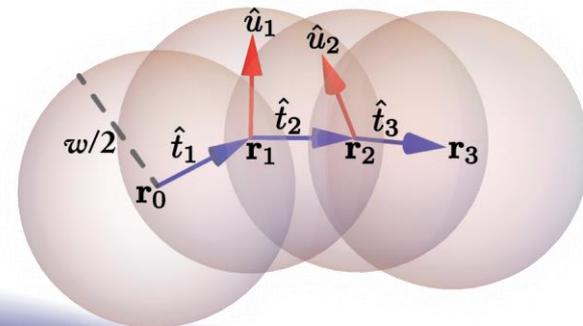
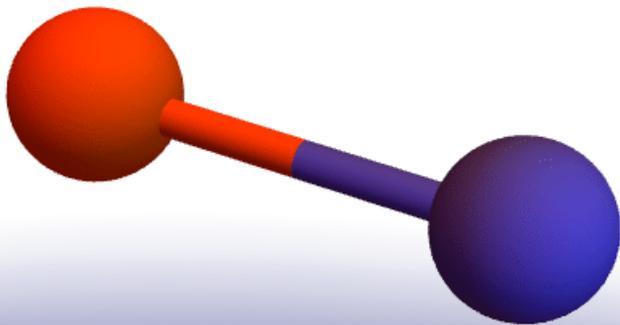
- Chains generated from scratch, link after link
- Links orientations mirror probabilities

• Samples of  $\sim 10^9$  chains with & without TFs

- Comparing looping probability with & without TFs determines regulatory effect.

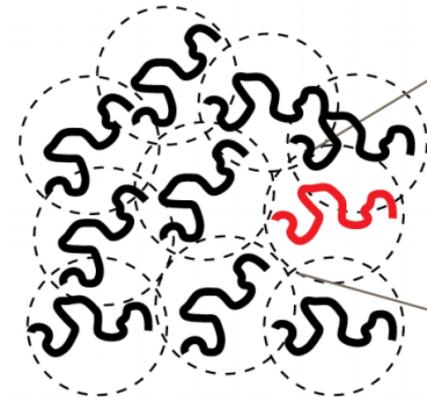
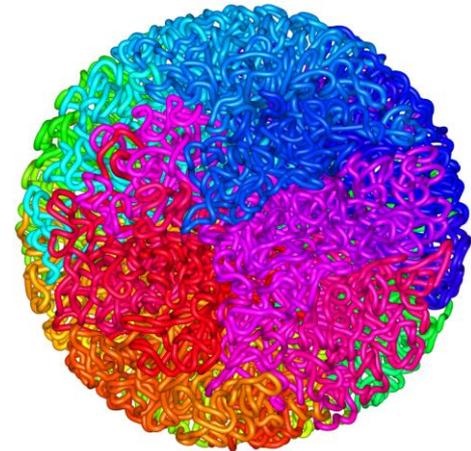
$$p_i(\{\theta_i, \phi_i\}) \propto \exp[-\beta E^{el+hw}(\{\theta_i, \phi_i\})] \Rightarrow P_{tot}(\{\theta_N, \phi_N\}) \propto \exp[-\beta E_{chain}^{el+hw}(\{\theta_N, \phi_N\})]$$

$$W(\{\theta_N, \phi_N\}) = \prod_{i=2}^N \left( \int_{-1}^1 d \cos \theta_i \int_0^{2\pi} d\phi_i \exp[-\beta E^{el}(\theta_i, \phi_i)] \Theta_i^{hw}(\{\theta_i, \phi_i\}) \right)$$



# Model Applicability

- Cellular DNA:
  - In condensed globular state
  - Divided into autonomic domains – “blobs”
  - Blob size varies 300-10<sup>3</sup> nm
- Linearized active enhancer-promoter regions in organisms with low chromatin volume fractions (yeast, *D. melanogaster*) can explore the volume of the blob without inter-chromatin interactions



# Confined DNA

- Confinement affects looping probability, not the ratio
- Enhancers/promoters are linearized
- Intermediate chain structure not important ( $ALA > b$ )
- Model applicable for small chromatin volume fractions

